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Iodonium Ion-Assisted Synthesis of Tetrameric Fragments Corresponding to the Cell Wall Phenolic Glycolipids of *Mycobacterium kansasii* serovars II and IV

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Abstract: The spacer-containing tetramers 3 and 4, derivatives of the phenolic glycolipids of *Mycobacterium kansasii* serovars II and IV were prepared by iodonium ion-mediated mannosylation of trimeric acceptor 4-[2-(benzyloxycarbonylamino)ethyl]phenyl 2,4-di-O-methyl-3-O-[4-O-benzyl-2-O-methyl-3-O-(4-O-acetyl-2-O-methyl-α-L-fucopyranosyl)-α-L-rhamnopyranosyl]-α-L-rhamnopyranoside (2) with ethyl 1-thio-D-mannopyranoside donors 7, 13, and 24. In addition, the glycosylating properties of donors 7, 13, 24, 29-31, each containing a different protective group at position 2, were examined by executing condensations with model acceptor 18(L) and its enantiomer 18(D).

INTRODUCTION

As part of a programme¹ to develop serodiagnostics and synthetic vaccines based on carbohydrates, we recently revealed² the assembly of the invariable $Mycobacterium\ kansasii$ inner core trimeric fragment 1, the terminal L-rhamnose unit of which is α -(O)-linked to a tyramine moiety suitable for conjugation with a protein.

We here report that glycosylation of the HO-3 in the L-fucopyranosyl unit of the partially protected trisaccharide acceptor 2 with appropriately protected ethyl 1-thio-D-mannopyranoside donors (i.e. 7, 13, and 24) gives access to the haptenic tetrameric fragments 3 and 4, which correspond to the linear oligosaccharides of M. kansasii phenolic glycolipid serovars II^3 and IV^4 , respectively.

- 1 R1=R2=R3=H
- 2 R1=Bn, R2=benzyloxycarbonyl(Z), R3=H

RESULTS AND DISCUSSION

The readily accessibility² of the partially protected trimeric fragment 2 prompted us to investigate whether 2 could be used as a starting compound for the construction of the target tetramers 3 and 4. A crucial step in the synthesis of tetrameric fragments 3 and 4 entails in both cases the stereoselective formation of a trans α -(1 \rightarrow 3)-interglycosidic bond between the HO-3 of the L-fucosyl unit in the trimeric acceptor 2 and an appropriately protected D-mannopyranosyl donor. However, the presence of an acetyl group at the O-4 position of the L-fucose residue in acceptor 2 restricts the use of mannopyranosyl donors bearing O-2 participating groups (*i.e.* acetyl or benzoyl). The limited choice in protecting groups imposed on the mannopyranosyl donors, and our broad experience in applying ethyl 1-thio-glycosides as building blocks, 1,2,5-8 were decisive factors in selecting ethyl 1-thio- α -D-mannopyranosides for the elongation of 2. Thus, condensation of acceptor 2 with ethyl 3,6-di-O-benzyl-2,4-di-O-methyl-1-thio- α -D-mannopyranoside (7) in the presence of the thiophilic promoter N-iodosuccinimide (NIS) and catalytic triflic acid (TfOH)⁵ would give the fully protected tetramer 21, a precursor of fragment 3. Similarly, glycosylation of 2 with ethyl 2,3,6-tri-O-benzyl-4-O-methyl-1-thio- α -D-mannopyranoside (13) would result in the formation of tetramer 22, a precursor of fragment 4.

Reagents and conditions: (i) Bu_2SnO , MeOH, 2.5 h; BnBr, CsF, DMF, 18 h, 59%. (ii) MeI, NaH, DMF, 1 h, 7 85%, 11 95%. (iii) TBAF, dioxane, 1.5 h, 79%. (iv) Bu_2SnO , MeOH, 1.5 h; BnBr, CsF, DMF, 18 h, 10 75%, 23 80%. (v) 90% HOAc, 50°C, 17 h, 89%. (vi) BnBr, NaH, DMF, 1.5 h, 88%. (vii) HCl-MeOH (1 N), reflux, 1.5 h; $CH_3C(OCH_3)_2$, acetone, TsOH, 1.5 h, $15-\alpha$ 42% and $15-\beta$ 27%. (viii) MeI, NaH, DMF, 1 h, 94%. (ix) 90% HOAc, 50°C, 17 h, 100%. (x) $CH_3C(OCH_3)_3$, CSA, CH_2CN , 45 min; 80% HOAc, 15 min, 81%. (xi) $(ClAc)_2O$, $NaHCO_3$, DMF, 24 h, 87%.

Scheme 1

The preparation of the required donors 7 and 13 starting from ethyl 1-thio- α -D-mannopyranoside⁸ (5) is depicted in Scheme 1. Thus, regioselective benzylation of the distannylidene complex⁹ of 5 with benzyl bromide, followed by methylation of the resulting 3,6-di-O-benzyl derivative 6, gave donor 7 in 50% overall yield. On the other hand, desilylation of known^{8b} ethyl 2,3-O-isopropylidene-6-O-(tert-butyldimethylsilyl)-1-thio- α -D-mannopyranoside (8) and regioselective benzylation¹⁰ of diol 9 gave the mono-benzylated derivative 10. Methylation of 10 and subsequent deacetonation of 11 afforded, after benzylation of diol 12, the partially benzylated donor 13 in 46% overall yield.

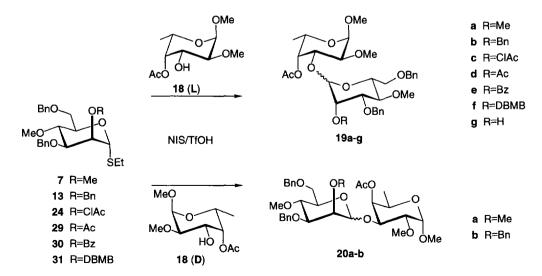
Prior to the intended elongation of trimeric fragment 2 with the mannopyranosyl donors 7 and 13, we first focused our attention on the stereochemistry of the glycosylation of these donors with the model compound methyl 2-O-methyl-4-O-acetyl- α -L-fucopyranoside [18(L)], prepared in four steps (see steps vii-x

Table 1 Relevant data on the NIS/TfOH(cat.)-assisted glycosylation* of fucopyranoside acceptor 18	(L) and
19(D) with D manuanyanasida danara 7, 12, 24, and 20, 21	

entry	donor	acceptor	dimer	yield (%)	α:β(:28)
1	7 R=Me	18(L)	19a	74	2:1
2	13 R=Bn	18(L)	19b	72	4:1
3	7 R=Me	18(D)	20a	68	4:1
4	13 R=Bn	18(D)	20b	65	6:1
5	24 R=ClAc	18(L)	19c, 28	78	6:1(:6)
6	24 R=ClAc**	18(L)	19c	79	6:1
7	29 R=Ac	18(L)	19d	63	1:0
8	30 R=Bz	18(L)	19e	92	1:0
9	31 R=DBMB	18 (L)	19f	94	1:0

^{*} Acceptor (1 equiv.), Donor (1.2 equiv.), NIS (1.2 equiv.), TfOH (0.1 equiv.), 1,2-dichloroethane-Et₂O, 0°C, 15 min.

in Scheme 1) from methyl $\alpha(\beta)$ -L-fucopyranoside¹¹ (14). The yield and stereochemical outcome of those pilot mannosylations are summarised in Table 1. It can be seen (Table 1, entries 1-2) that NIS/TfOH(cat.)-assisted glycosylations of acceptor 18(L) with 2-O-Me- and 2-O-Bn-mannopyranosides 7 and 13 resulted in the formation of a high percentage β -linked dimers 19a and 19b (Scheme 2). The unfavourable outcome of the latter glycosylations might be ascribed to the occurrence of double stereodifferentiation¹² (DSD). In order to validate this assumption, the enantiomeric acceptor 18(D), prepared in three steps (*i.e.* steps viii-x in Scheme 1) from 15(D)¹³, was also condensed with the same donors 7 and 13. Iodonium ion-mediated glycosylation of 18(D) with mannopyranosyl donors 7 and 13 gave the respective dimers 20a and 20b as a mixture of anomers (Table 1, entries 3-4). A comparison between the stereochemical outcome of the latter glycosylations demonstrates that different α : β ratios are observed in the coupling of the individual enantiomers of 18 with both donors 7 (R=Me) and 13 (R=Bn), indicating that the occurrence of DSD, albeit to a minor degree, is not excluded.



Scheme 2

^{** 0.3} equiv. TfOH was used.

In the light of the aforementioned pilot glycosylations, it was anticipated that the stereochemistry of the condensation of the L-trimeric acceptor 2 with the D-mannopyranosyl donors 7 and 13 would not deviate substantially from those of the same donors with the L-acceptor 18 (see Table 1, entries 1-2). Indeed, NIS/TfOH(cat.)-promoted D-mannosylation of 2 with 7 (see Scheme 3) gave tetramer 21 as a mixture of anomers (α : β =3:1). Separation of the individual anomers by silica gel chromatography afforded the requisite α -mannosylated tetramer 21 in 63% yield. Similar to the synthesis of 21, purification of the crude mannosylation mixture (α : β =4:1) resulting from the elongation of 2 with 13 gave the α -mannosylated tetramer 22 in 60% yield. Removal of the benzyl (Bn) and benzyloxycarbonyl (Z) groups from both tetramers 21 and 22 led, after purification by Sephadex LH20 gel-filtration, to the isolation of the respective target tetramers 3 and 4, the structures of which were firmly established by NMR-spectroscopy.

NHR¹

BnO

OMe

$$2 + 7 (13, 24)$$

ii

 $-21 R^1 = Z, R^2 = Bn, R^3 = Me; \alpha : \beta = 3:1$

3 $R^1 = R^2 = H, R^3 = Me$

ii

 $-22 R^1 = Z, R^2 = R^3 = Bn; \alpha : \beta = 4:1$

4 $R^1 = R^2 = R^3 = H$

iii

 $-25 R^1 = Z, R^2 = Bn, R^3 = CIAc$

iii

 $-25 R^1 = Z, R^2 = Bn, R^3 = CIAc$

iii

 $-27 R^1 = Z, R^2 = Bn, R^3 = OH$

Reagents and conditions: (i) NIS/TfOH(cat.), 1,2-dichloroethane-Et₂O, 0°C, 15 min, 21 83%, 22 74%, 25+26 76%. (ii) H₂, Pd(C), isopropanol-H₂O-HOAc, 66 h, 3 86%, 4 86%, 85%. (iii) HDTC, HOAc, lutidine, 0°C, 30 min, 86%.

Scheme 3

At this stage, we were interested to find out whether the rather low α -stereoselectivity in the mannosylation of L-acceptor 2 with the D-donors 7 and 13 could be increased by the known¹⁴ α -directing nature of the participating medium diethyl ether. Pilot experiments indicated that α -mannosylation of 18(L) by 7 at 20°C in diethyl ether proceeded with a high degree of stereoselectivity to give dimer 19a as a mixture of anomers in the ratio α : β =7:1. As expected, the latter beneficial stereochemical effect was also observed in the glycosylation of trimeric acceptor 2 with both donors 7 and 13. Thus, the individual tetrameric fragments 21 and 22 were obtained as an anomeric mixture (α : β ratio 10:1 and 12:1, respectively) and in excellent yield (\approx 90%).

It was expected that the formation of the unwanted β -anomer could be suppressed by replacing the non-participating 2-O-benzyl by a participating chloroacetyl group, the removal of which is compatible with the presence of a 4-O-acetyl group in the fucosyl moiety.

The requisite donor ethyl 3,6-di-*O*-benzyl-2-*O*-chloroacetyl-4-*O*-methyl-1-thio-α-D-mannopyranoside (24 in Scheme 1) was obtained in 69% overall yield via regioselective benzylation of the equatorial hydroxyl group in 12 and subsequent chloroacetylation of 23. However, glycosylation of 2 with the 2-*O*-chloroacetyl-D-mannopyranosyl donor 24 in the presence of NIS and variable amounts of TfOH (0.1 or 0.3 equiv.) gave tetrameric fragment 25 (Scheme 3) and its 1,2-orthoester derivative 26 (see Figure 3), as evidenced by NMR spectroscopy¹⁵. Unfortunately, every attempt to convert 26 into 25 via acid-catalysed (TfOH) rearrangement¹⁶ was accompanied by the formation of the trimeric fragment fragment 2. Selective removal of the chloroacetyl group in 25 with hydrazine dithiocarbonate¹⁷ (HDTC) and subsequent hydrogenolysis of resulting 27 gave, after purification by Sephadex LH20 gel-filtration, homogeneous 4, which was in every aspect identical with the same tetrasaccharide obtained by hydrogenolysis of 22.

In this respect it is of interest to note that mannosylation (Table 1, entry 5) of 18(L) with donor 24, having a participating 2-O-chloroacetyl group, led to an anomeric mixture of dimer 19c (Scheme 2) and its 1,2-orthoester derivative 28. The identity of compound 28 was ascertained by 1H - and ^{13}C -NMR spectroscopy 15 and its quantitative acid-catalysed (TfOH) rearrangement 16 into the α -linked dimer of 19c (cf. conversion $26 \rightarrow 25$). Apart from this, it was established that the formation of the rather acid-stable orthoester 28 could be prevented by executing the NIS-mediated mannosylation in the presence of relative excess triflic acid (see entry 6 in Table 1). The formation of the undesired β -dimer of 19c in the latter two mannosylations may be due to the less effective participating aptitude of the 2-O-chloroacetyl group. Indeed, exclusive α -mannosylation of 18(L) occurred (Table 1, entries 7-8) using the corresponding 2-O-acetyl (29) or benzoyl (30) derivatives, obtained by acetylation or benzoylation of 23, as the glycosylating agents.

The stereoselective introduction of the 1,2-trans linkage between 2-O-benzoyl-D-mannopyranoside 30 and the L-fucopyranoside acceptor 18 urged us to explore whether the D-mannopyranoside 31, in which HO-2 is protected with the versatile 2-dibromomethylbenzoyl (DBMB) group¹⁹, is a viable building unit in the synthesis of the partially protected dimer 19g (see Scheme 2). The requisite mannopyranoside donor 31 was readily accessible by acylation of 23 with 2-dibromomethylbenzoyl chloride. Iodonium ion-mediated glycosylation of the L-fucosyl acceptor 18 with 31 proceeded in a stereoselective manner to give the α-linked disaccharide 19f in an excellent yield (see entry 9 in Table 1). Treatment of 31 with silver perchlorate in acetone-water in the presence of 2,6-lutidine, and subsequent addition of morpholine to the resulting 2-O-(2-formyl)benzoyl derivative, gave partially protected dimer 19g in 84% over the two steps. The successful synthesis of 19g clearly demonstrates that the two-step removal of the DBMB is compatible with the presence of the 4-O-acetyl in the fucosyl moiety. It may therefore not be excluded that the DBMB group presents an attractive alternative for the recently proposed²⁰ 2-(2-chloroacetoxyethyl)benzoyl group, which is cleavable besides other acyl groups via a rather sluggish two-step process.

EXPERIMENTAL

General methods and materials: Methanol was dried by refluxing with magnesium methoxide and then distilled. Toluene and 1,2-dichloroethane were distilled from P_2O_5 . N,N-Dimethylformamide (DMF) was stirred with calcium hydride for 20 h, then distilled under reduced pressure. Pyridine and dioxane were dried by refluxing 18 h with calcium hydride and then distilled. Acetonitrile (p.a. Rathburne) was dried by storage over molecular sieves 4 Å (Aldrich). Diethyl ether was distilled from LiAlH₄. Methanol was stored over molecular sieves 3 Å. Toluene and diethyl ether were stored over sodium wire. DMF, dioxane, pyridine and 1,2-dichloroethane were stored over molecular sieves 4 Å.

Reactions were performed under anhydrous conditions at room temperature unless stated otherwise. Evaporation of solvents was performed under reduced pressure at 40°C. TLC-analyses were conducted on DC Fertigfolien (Schleicher & Schüll

F 1500, LS 254). Compounds were visualised with UV light (254 nm) and by charring with concentrated sulfuric acid/ethanol (1/4, v/v). Column chromatography was performed on columns of silica gel 60, 230-400 mesh (Merck). The petroleum ether used for eluting the columns was low boiling (40-60°C). Sephadex LH20 (Pharmacia) was used for gel-filtration.

Optical rotations were determined with a Propol polarimeter for solutions in CHCl₃ (p.a. Baker) at 20°C. ¹H-NMR (200 MHz) and ¹³C-NMR (50.1 MHz) spectra were recorded using a Jeol JNM-FX-200 spectrometer. Spectra were also recorded using a Bruker WM-300 spectrometer equipped with an Aspect 2000 computer, and a Bruker MSL-400 connected with an Aspect 3000 computer. Chemical shifts are given in ppm (δ) relative to tetramethylsilane as an internal standard. Mass spectra of compounds dissolved in methanol-water (4/1, v/v) were recorded with a Finnigan MAT TSQ-70 equipped with a custom-made Electrospray Interface (ESI).

Ethyl 3,6-di-O-benzyl-1-thio- α -D-mannopyranoside (6) - To a solution of ethyl 1-thio- α -D-mannopyranoside (5, 1.13 g, 5.0 mmol) in methanol (41 ml) was added dibutyltin oxide (2.65 g, 10.6 mmol). The mixture was heated under reflux for 2.5 h. The solvent was removed and the residue was dried by evaporation with toluene. The distannylidene derivative was dissolved in DMF (34 ml), cesium fluoride (2.14 g, 14.1 mmol) and benzyl bromide (1.70 ml, 14.4 mmol) were added and the reaction mixture was stirred for 18 h at room temperature. The solvent was evaporated and the residue was redissolved in diethyl ether (20 ml). The organic layer was washed twice with aq. KF (1 M, 15 ml) and once with H_2O (10 ml), dried (MgSO₄), filtered, and concentrated. The oily residue was purified by column chromatography (ethyl acetate in petroleum ether $O \rightarrow 30\%$) to furnish compound 6 (1.20 g, 3.0 mmol).

¹³C{¹H}-NMR (CDCl₃): δ 14.9 (CH₃ SEt), 24.9 (CH₂ SEt), 70.0 (C-6), 71.7, 73.4 (2× CH₂ Bn), 67.4, 69.4, 71.8, 79.9 (C-2, C-3, C-4, C-5), 84.0 (C-1), 126.9, 127.2, 127.6, 127.9, 128.1, 128.3, 128.5 (CH Bn), 137.9, 138.3 (qC Bn). Anal. calcd. for $C_{22}H_{28}O_3S$ (404.53): C 65.32, H 6.98; found C 65.44, H 7.06%.

Ethyl 3,6-di-O-benzyl-2,4-di-O-methyl-1-thio- α -D-mannopyranoside (7) - Methyl iodide (0.49 ml, 7.8 mmol) was added at 0°C to a suspension of compound 6 (1.20 g, 3.0 mmol) and sodium hydride (60%, 360 mg, 9.0 mmol) in DMF (6 ml). After stirring for 45 min at room temperature, the reaction was quenched with methanol (1 ml) and the solvents were evaporated. A solution of the residue in diethyl ether (20 ml) was washed with H_2O (15 ml) and aq. NaHCO₃ (10%, 15 ml), dried (MgSO₄), filtered, and concentrated. Purification of the oily residue by column chromatography (0 \rightarrow 10% ethyl acetate in petroleum ether) gave mannopyranoside donor 7 (1.09 g, 2.5 mmol).

¹H-NMR (CDCl₃): δ 1.27 (t, 3H, CH₃ SEt, $J_{\text{H,H}}$ 7.4 Hz), 2.61 (AB, 2H, CH₂ SEt), 3.44, 3.49 (2× s, 6H, 2× CH₃ Me), 3.52 (dd, 1H, H-2, $J_{2,1}$ 1.5 Hz, $J_{2,3}$ 3.1 Hz), 3.61-3.80 (m, 3H, H-6, H-3), 4.01 (ddd, 1H, H-5, $J_{5,4}$ 9.2 Hz, $J_{5,6}$ 4.6 Hz, $J_{5,6}$ 2.0 Hz), 4.48-4.72 (m, 4H, 2× CH₂ Bn), 5.40 (d, 1H, H-1, $J_{1,2}$ 1.5 Hz), 7.31-7.40 (m, 10H, CH Bn); ¹³C(¹H}-NMR (CDCl₃): δ 14.9 (CH₃ SEt), 25.1 (CH₂ SEt), 58.2, 60.6 (2× CH₃ Me), 69.2 (C-6), 72.1, 73.1 (2× CH₂ Bn), 71.9, 76.5, 79.4, 80.1 (C-2, C-3, C-4, C-5), 81.0 (C-1), 127.3, 127.5, 127.7, 128.1, 128.2 (CH Bn), 138.3, 138.4 (qC Bn).

Anal. calcd. for C₂₄H₃₂O₅S (432.58): C 66.64, H 7.46; found C 66.53, H 7.39%.

Ethyl 2,3-O-isopropylidene-1-thio- α -D-mannopyranoside (9) - Compound 8 (3.87 g, 11.2 mmol) was dried by repeated evaporation with dioxane and dissolved in the same solvent (37 ml). A solution of tetrabutylammonium fluoride (TBAF) in dioxane (1 M, 20 ml) was added and the mixture was stirred for 1.5 h. After concentration of the reaction mixture, the crude product was purified by column chromatography. The column was eluted with $40\rightarrow60\%$ ethyl acetate in petroleum ether. Concentration of the appropriate fractions yielded compound 9 (2.33 g, 8.8 mmol).

 13 C{ 1 H}-NMR (CDCl₃): δ 14.4 (CH₃ SEt), 24.1 (CH₂ SEt), 26.2, 28.0 (2× CH₃ Isopr), 61.7 (C-6), 69.4, 69.7, 76.4, 78.3 (C-2, C-3, C-4, C-5), 79.5 (C-1), 109.5 (qC Isopr).

Ethyl 6-O-benzyl-2,3-O-isopropylidene-1-thio-α-p-mannopyranoside (10) - To a solution of compound 9 (845 mg, 3.2 mmol) in methanol (20 ml) was added dibutyltin oxide (876 mg, 3.5 mmol). The mixture was heated under reflux for 1 h, and concentrated. The residue was dried by repeated evaporation with toluene and redissolved in DMF (20 ml). Benzyl bromide (0.57 ml, 4.8 mmol) and cesium fluoride (634 mg, 4.2 mmol) were added. After stirring for 18 h, DMF was removed and the residue was taken up in diethyl ether (25 ml). The solution was washed twice with aq. KF (1 M, 15 ml), once with water (15 ml), dried (MgSO₄), filtered, and the solvent was evaporated. The crude product was purified by silica gel column chromatography. Elution

of the column with $0\rightarrow60\%$ ethyl acetate in petroleum ether gave 10 (749 mg, 2.4 mmol) and starting compound 9 (85 mg, 0.3 mmol).

9: ¹³C{¹H}-NMR (CDCl₃): δ 14.3 (CH₃ SEt), 23.9 (CH₂ SEt), 26.2, 28.0 (2× CH₃ Isopr), 69.6 (C-6), 73.3 (CH₂ Bn), 69.4, 70.1, 76.3, 78.3 (C-2, C-3, C-4, C-5), 79.2 (C-1), 109.4 (qC Isopr), 126.8, 127.4, 128.2 (CH Bn), 138.1 (qC Bn).

Ethyl 6-O-benzyl-4-O-methyl-1-thio- α -D-mannopyranoside (12) - Compound 10 (749 mg, 2.4 mmol) was dissolved in DMF (4 ml) and sodium hydride (60%, 145 mg, 3.6 mmol) and methyl iodide (0.19 ml, 3.1 mmol) were added at 0°C. After stirring for 1 h at room temperature, the reaction was quenched with methanol (2 ml), and the solvents were removed. The residue was taken up in diethyl ether (15 ml). The solution was washed with water (10 ml) and aq. NaHCO₃ (10%, 10 ml), dried (MgSO₄), filtered, and concentrated. The thus obtained fully protected mannopyranoside 11 was dissolved in acetic acid-water (9/1, v/v, 18 ml) and the mixture was heated at 50°C for 17 h. The solution was concentrated and the residue was evaporated with toluene. The residue was purified by a silica gel column chromatography. Elution with ethyl acetate in petroleum ether (0 \rightarrow 60%) yielded compound 12 (701 mg, 2.1 mmol).

¹³C{¹H}-NMR (CDCl₃): δ 14.7 (CH₃ SEt), 24.6 (CH₂ SEt), 60.2 (CH₃ Me), 69.1 (C-6), 73.1 (CH₂ Bn), 71.0, 72.0, 72.3, 77.4 (C-2, C-3, C-4, C-5), 84.1 (C-1), 127.3, 127.5, 128.1 (CH Bn), 138.0 (qC Bn).

Anal. calcd. for C₁₆H₂₄O₅S (328.43): C 58.51, H 7.37; found C 58.59, H 7.32%.

Ethyl 2,3,6-tri-O-benzyl-4-O-methyl-1-thio- α -D-mannopyranoside (13) - To a cooled (0°C) solution of compound 12 (313 mg, 1.0 mmol) in DMF (2 ml) were added sodium hydride (60%, 120 mg, 3.0 mmol) and benzyl bromide (0.31 ml, 2.6 mmol). After stirring for 1.5 h, the reaction was quenched with methanol (1 ml) and the solvents were evaporated. The residue was redissolved in diethyl ether (15 ml), and the solution was washed with water (10 ml) and aq. NaHCO₃ (10%, 10 ml), dried (MgSO₄), and filtered. The filtrate was concentrated and the residual oil was purified by column chromatography. The column was eluted with ethyl acetate in petroleum ether (0 \rightarrow 5%) to give pure 13 (471 mg, 0.9 mmol).

[α]_p +103.0° (c 1); ¹³C{¹H}-NMR (CDCl₃): δ 15.0 (CH₃ SEt), 25.2 (CH₂ SEt), 60.8 (CH₃ Me), 69.3 (C-6), 71.9, 72.0, 73.3 (3× CH₂ Bn), 72.2, 76.5, 76.7, 80.3 (C-2, C-3, C-4, C-5), 81.8 (C-1), 127.3, 127.5, 127.8, 128.3 (CH Bn), 138.2, 138.4, 138.5 (qC Bn).

Anal. calcd. for C₃₀H₃₆O₅S (508.68): C 70.82, H 7.13; found C 70.90, H 7.06%.

Methyl 3,4-O-isopropylidene-α-L-fucopyranoside [15(L)] - L-Fucopyranose (5.10 g, 31.1 mmol) was treated with HCl in methanol (0.5 M, 100 ml), according to Schuler et al. ¹⁴ to give, after neutralisation of the reaction mixture and evaporation of the solvent, methyl pyranoside 14(L).

Compound 14(L) was dissolved in a mixture of acetone (30 ml) and dimethoxypropane (16 ml). p-Toluenesulfonic acid (571 mg, 3 mmol) was added and the reaction mixture was stirred for 2 h. The solution was neutralised with Et₃N and the solvents were evaporated. The residue was taken up in ethyl acetate (150 ml) and the solution was washed with water (100 ml) and aq. NaHCO₃ (10%, 100 ml), dried (MgSO₄), and filtered. Ethyl acetate was evaporated and the oily residue was purified by column chromatography. The column was eluted with 0 \rightarrow 40% ethyl acetate in petroleum ether to give the α -fucopyranoside 15(L)- α (2.82 g, 12.9 mmol) and the β -linked anomer 15(L)- β (1.71 g, 7.8 mmol).

15(L)-α: ¹H-NMR (CDCl₃): δ 1.33 (d, 3H, H-6, J_{6.5} 6.7 Hz), 1.36, 1.52 (2× s, 6H, 2× CH₃ Isopr), 2.28 (d, 1H, OH, J_{HO.2} 6.7 Hz), 3.44 (s, 3H, CH₃ 1-*O*-Me), 3.79 (dt, 1H, H-2, J_{2,1} 3.9 Hz, J_{2,3}≈J_{2,HO} 6.7 Hz), 4.01-4.13 (m, 2H, H-3, H-4), 4.18 (q, 1H, H-5, J_{5.6} 6.2 Hz), 4.72 (d, 1H, H-1, J_{1,2} 3.9 Hz); ¹³C{¹H}-NMR (CDCl₃): δ 16.2 (C-6), 25.9, 27.8 (2× CH₃ Isopr), 55.3 (CH₃ 1-*O*-Me), 63.5, 69.4, 75.6, 76.2 (C-2, C-3, C-4, C-5), 98.7 (C-1), 109.0 (qC Isopr).

15(L)-β: 1 H-NMR (CDCl₃): δ 1.36 (s, 3H, CH₃ Isopr), 1.43 (d, 3H, H-6, J_{6.5} 6.7 Hz), 1.53 (s, 3H, CH₃ Isopr), 2.47 (d, 1H, OH, J_{HO.2} 2.3 Hz), 3.48 (dd, 1H, H-3, J_{3.2} 8.4 Hz, J_{3.4} 5.5 Hz), 3.54 (s, 3H, CH₃ 1-*O*-Me), 3.87 (dq, 1H, H-5, J_{5.4} 2.1 Hz, J_{5.6} 6.6 Hz), 4.01-4.10 (m, 3H, H-1, H-2, H-4); 13 C{ 1 H}-NMR (CDCl₃): δ 16.4 (C-6), 26.2, 28.1 (2× CH₃ Isopr), 56.7 (CH₃ 1-*O*-Me), 68.9, 73.4, 76.2, 78.8 (C-2, C-3, C-4, C-5), 103.1 (C-1), 109.6 (qC Isopr).

Methyl 3,4-O-isopropylidene-2-O-methyl-α-L-fucopyranoside [16(L)] - Compound 15 was (1.84 g, 8.4 mmol) treated with sodium hydride (60%, 504 mg, 12.6 mmol) and methyl iodide (0.68 ml, 10.9 mmol) in DMF (16 ml), as described for the preparation of compound 12, to give crude 16(L) (1.83 g, 7.8 mmol).

 1 H-NMR (CDCl₃): δ 1.36 (d, 3H, H-6, J_{6,5} 6.7 Hz), 1.36, 1.55 (2× s, 6H, 2× CH₃ Isopr), 3.36 (dd, 1H, H-3, J_{3,2} 7.8 Hz), 3.41, 3.53 (2× s, 6H, 2× CH₃ Me), 4.02-4.14 (m, 2H, H-4, H-5), 4.23 (br dd, 1H, H-2, J_{2,1} 5.4 Hz, J_{2,3} 7.7 Hz), 4.79 (d, 1H, H-1, J_{2,1} 3.6 Hz); 13 C{ 1 H}-NMR (CDCl₃): δ 15.8 (C-6), 25.8, 27.9 (2× CH₃ Isopr), 54.8 (CH₃ 1-*O*-Me), 58.1 (CH₃ Me), 62.4 (C-5), 75.4, 75.6, 78.8 (C-2, C-3, C-4), 97.4 (C-1), 108.4 (qC Isopr).

Methyl 2-O-methyl-α-L-fucopyranoside [17(L)] - Crude 16(L) (1.83 g, 7.8 mmol) was dissolved in acetic acid-water (9/1, v/v, 5 ml) and stirred for 17 h at 50°C. Concentration of the solution followed by evaporation of the residue with toluene gave 17(L) (1.47 g, 7.8 mmol) which was used without further purification.

 1 H-NMR (CDCl₃); δ 1.30 (d, 3H, H-6, $J_{6.5}$ 6.7 Hz), 3.16 (s, 2H, 2× OH), 3.41, 3.49 (2× s, 6H, 2× CH₃ Me), 3.52 (dd, 1H, H-3, $J_{3.2}$ 9.8 Hz, $J_{3.4}$ 3.6 Hz), 3.80 (d, 1H, H-4, $J_{4.3}$ 3.3 Hz), 3.84-3.97 (m, 2H, H-2, H-5, $J_{2.3}$ 9.8 Hz, $J_{2.1}$ 3.1 Hz, $J_{5.6}$ 6.3 Hz), 4.90 (d, 1H, H-1, $J_{1.2}$ 3.6 Hz); 13 C[1 H}-NMR (CDCl₃): δ 15.9 (C-6), 54.7 (CH₃ 1-*O*-Me), 57.5 (CH₃ Me), 65.3 (C-5), 69.3, 71.7, 77.6 (C-2, C-3, C-4), 97.0 (C-1).

Methyl 4-O-acetyl-2-O-methyl-α-L-fucopyranoside [18(L)] - To a solution of compound 17(L) (1.47 g, 7.8 mmol) in acetonitrile (23 ml) were added trimethyl orthoacetate (2.0 ml, 15.6 mmol) and camphorsulfonic acid (182 mg, 0.78 mmol). After stirring for 45 min, a mixture of acetic acid-water (4/1, v/v, 45 ml) was added and stirring was continued for 15 min. The solution was diluted with dichloromethane (50 ml) and the layers were separated. The organic layer was washed with H_2O (30 ml) and aq. NaHCO₃ (10%, 30 ml), dried (MgSO₄), and filtered. The organic solvents were removed and the oily residue was purified by column chromatography (ethyl acetate in petroleum ether, 20 \rightarrow 60%) to yield 18(L) (1.48 g, 6.4 mmol).

[α]_D -168.6° (c 1); ¹H-NMR (CDCl₃): δ 1.16 (d, 3H, H-6, J_{6.5} 6.7 Hz), 2.19 (s, 3H, CH₃ Ac), 2.42 (d, 1H, OH, J_{HO,3} 2.8 Hz), 3.43 (dd, 1H, H-2/H-3, J_{H,H} 3.6 Hz, J_{H,H} 9.8 Hz), 3.43, 3.53 (2× s, 6H, 2× CH₃ Me), 4.04 (q, 1H, H-5, J_{6.5} 6.7 Hz), 4.08-4.11 (m, 1H, H-2/H-3), 4.92 (d, 1H, H-1/H-4 J_{H,H} 3.3 Hz), 5.24 (d, 1H, H-1/H-4, J_{H,H} 3.6 Hz); ¹³C{¹H}-NMR (CDCl₃): δ 15.6 (C-6), 20.3 (CH₃ Ac), 54.9 (CH₃ 1-*O*-Me), 57.8 (CH₃ Me), 64.1 (C-5), 67.3, 73.0, 77.7 (C-2, C-3, C-4), 97.0 (C-1), 170.7 (C=O Ac).

Anal. calcd. for C₁₀H₁₈O₆ (234.25): C 51.27, H 7.75; found C 51.38, H 7.83%.

Methyl 3,4-O-isopropylidene-2-O-methyl-α-D-fucopyranoside [16(D)] - Known¹⁰ methyl 3,4-O-isopropylidene-α-D-fucopyranoside [15(D), 0.90 g, 4.1 mmol] was treated with sodium hydride and methyl iodide, as described for the preparation of 12, to yield compound 16(D) (0.90 g, 3.9 mmol).

 1 H-NMR (CDCl $_{3}$): δ 1.36 (d, 3H, H-6, J $_{6,5}$ 6.4 Hz), 1.36, 1.55 (2× s, 6H, 2× CH $_{3}$ Isopr), 3.36 (dd, 1H, H-2, J $_{2,1}$ 3.5 Hz, J $_{2,3}$ 7.8 Hz), 3.41, 3.49 (2× s, 6H, 2× CH $_{3}$ Me), 4.01-4.05 (m, 1H, H-4), 4.11 (dq, 1H, H-5, J $_{5,4}$ 2.4 Hz, J $_{5,6}$ 6.6 Hz), 4.23 (dd, 1H, H-3, J $_{3,2}$ 7.8 Hz, J $_{3,4}$ 5.2 Hz), 4.79 (d, 1H, H-1, J $_{1,2}$ 3.4 Hz); 13 C{ 1 H}-NMR (CDCl $_{3}$): δ 15.9 (C-6), 25.9, 27.9 (2× CH $_{3}$ Isopr), 54.9 (CH $_{3}$ 1-O-Me), 58.2 (CH $_{3}$ Me), 62.5, 75.5, 75.7, 78.9 (C-2, C-3, C-4, C-5), 97.4 (C-1), 108.2 (qC Isopr).

Methyl 2-O-methyl- α -D-fucopyranoside [17(D)] - Crude 16(D) (0.90 g, 3.9 mmol) was dissolved in acetic acid-water (9/1, v/v, 20 ml) and stirred for 18 h at 50°C. The solvents were evaporated under reduced pressure and the residual acetic acid was removed by repeated evaporation with toluene to afford compound 17(D) (0.75 g, 3.9 mmol).

 1 H-NMR (CDCl₃): δ 1.31 (d, 3H, H-6, J_{6,5} 6.9 Hz), 2.46 (br s, 1H, OH), 2.69 (br s, 1H, OH), 3.40-3.49 (m, 2H, H-2, H-3), 3.43, 3.49 (2× s, 6H, 2× CH₃ Me), 3.59 (dd, 1H, H-4, J_{4,3} 3.4 Hz, J_{4,5} 1.7 Hz), 3.94 (br q, 1H, H-5, J_{5,6} 6.6 Hz), 4.01-4.05 (m, 1H, H-4), 4.23 (dd, 1H, H-3, J_{3,2} 7.8 Hz, J_{3,4} 5.2 Hz), 4.91 (d, 1H, H-1, J_{1,2} 3.4 Hz); 13 C{ 1 H}-NMR (CDCl₃): δ 15.9 (C-6), 25.9, 27.9 (2× CH₃ Isopr), 54.9 (CH₃ 1-*O*-Me), 58.2 (CH₃ Me), 62.5, 75.5, 75.7, 78.9 (C-2, C-3, C-4, C-5), 97.4 (C-1), 108.2 (qC Isopr).

Methyl 4-O-acetyl-2-O-methyl-α-D-fucopyranoside [18(D)] - Crude 17(D) (0.75 g, 3.9 mmol) was converted in compound 18(D) (0.73 g, 3.1 mmol) as described for the enantiomer 18(L).

[α]_D +137.2°; ¹H-NMR (CDCl₃): δ 1.16 (d, 3H, H-6, J_{6,5} 6.6 Hz), 2.19 (s, 3H, CH₃ Ac), 2.28 (d, 1H, OH, J_{H0,2} 2.8 Hz), 3.43 (s, 3H, CH₃ Me), 3.49 (dd, 1H, H-2/H-3, J_{HH} 3.4 Hz), 3.51 (s, 3H, CH₃ Me), 4.03 (q, 1H, H-5, J_{5,6} 6.6 Hz), 4.07-4.14 (m, 1H, H-2/H-3), 4.92 (d, 1H, H-1/H-4, J_{HH} 3.6 Hz), 5.23 (d, 1H, H-1/H-4, J_{HH} 3.6 Hz); ¹³C{¹H}-NMR (CDCl₃): δ

15.5 (C-6), 20.1 (CH₃ Ac), 54.6 (CH₃ 1-*O*-Me), 57.6 (CH₃ Me), 64.0, 67.2, 73.0, 77.6 (C-2, C-3, C-4, C-5), 96.9 (C-1), 170.4 (C=O Ac).

Anal. calcd. for C₁₀H₁₈O₆ (234.25): C 51.27, H 7.75; found C 51.35, H 7.69%.

General glycosylation of the fucosyl acceptors 18(L) or 18(D)

A solution of glycosyl donor (0.3 mmol) and fucosyl acceptor 18 (58 mg, 0.25 mmol) in 1,2-dichloroethane-diethyl ether (1/1, v/v, 2 ml) was stirred in the presence of activated molecular sieves (4 Å) for 30 min. The mixture was cooled (0°C), and a suspension of NIS (68 mg, 0.3 mmol) and TfOH (2.8 μ l, 32 μ mol) in the same solvent mixture (2 ml) was added. After stirring for 15 min, the reaction was quenched with pyridine, filtered, and diluted with ethyl acetate (15 ml). The solution was washed with aq. Na₂S₂O₃ (20%, 10 ml) and aq. NaHCO₃ (10%, 10 ml), dried (MgSO₄), filtered, and concentrated. The reaction mixture was purified by silica gel column chromatography. The column was eluted with ethyl acetate in petroleum ether (0 \rightarrow 40%) to give the individual pure dimers (unless stated otherwise).

Methyl 4-O-acetyl-3-O-(3,6-di-O-benzyl-2,4-di-O-methyl- α -D-mannopyranosyl)-2-O-methyl- α -L-fucopyranoside (19a- α) and Methyl 4-O-acetyl-3-O-(3,6-di-O-benzyl-2,4-di-O-methyl- β -D-mannopyranosyl)-2-O-methyl- α -L-fucopyranoside (19a- β) - Glycosylation of acceptor 18(L) with donor 7 gave, after column chromatography, the α -linked dimer 19a- α (86 mg, 0.11 mmol) and by the β -linked isomer 19a- β (45 mg, 0.07 mmol).

The glycosylation of acceptor 18(L) with donor 7 was also performed in diethyl ether at room temperature. After stirring donor and acceptor in the presence of molecular sieves (4 Å) for 30 min, NIS was added followed by a solution of TfOH in diethyl ether. The reaction was further processed as described above. Purification by column chromatography gave an anomeric mixture of 19a (136 mg, 0.23 mmol) in the ratio α : β =7:1, based on the ¹H-NMR spectrum.

19a-α: [α]_b -21.7° (c 1); ¹H-NMR (CDCl₃, 300 MHz, HH-COSY): δ 1.03 (d, 3H, H-6, $J_{6.5}$ 6.5 Hz), 2.04 (s, 3H, CH₃ Ac), 3.40, 3.44 (2× s, 6H, 2× CH₃ Me), 3.42 (dd, 1H, H-2', $J_{2,1}$ 1.6 Hz, $J_{2,3}$ 3.4 Hz), 3.43, 3.49 (2× s, 6H, 2× CH₃ Me), 3.49 (dd, 1H, H-2, $J_{2,1}$ 3.6 Hz, $J_{2,3}$ 10.2 Hz), 3.60 (dd, 1H, H-3', $J_{3,2}$ 3.2 Hz, $J_{3,4}$ 9.2 Hz), 3.58 (t, 1H, H-4', $J_{4,3} \approx J_{4,5}$ 9.2 Hz), 3.56-3.85 (m, 3H, H-5', H-6'), 3.88 (dq, 1H, H-5, $J_{5,4}$ 1.2 Hz, $J_{5,6}$ 6.5 Hz), 4.09 (dd, 1H, H-3, $J_{3,2}$ 10.2 Hz, $J_{3,4}$ 3.5 Hz), 4.61, 4.72 (2× AB, 4H, 2× CH₂ Bn), 4.85 (d, 1H, H-1, $J_{1,2}$ 3.6 Hz), 5.14 (d, 1H, H-1', $J_{1,2}$ 1.8 Hz), 5.17 (dd, 1H, H-4, $J_{4,3}$ 3.5 Hz, $J_{4,5}$ 1.2 Hz), 7.15-7.43 (m, 10H, CH Bn); ¹³C{¹H}-NMR (CDCl₃): δ 15.7 (C-6), 20.4 (CH₃ Ac), 55.1, 58.4, 60.2 (3× CH₃ Me), 64.5 (C-5), 69.0 (C-6'), 71.9, 72.8 (2× CH₂ Bn), 71.9, 73.0, 76.3, 78.2, 78.5 (CH sugar rings), 97.3, 98.5 (C-1, C-1', $^{1}J_{CH}$ 171.4, 170.0 Hz, respectively), 127.0, 127.3, 127.4, 127.9, 128.0, 128.7 (CH Bn), 138.5 (qC Bn), 170.4 (C=O Ac).

Anal. calcd. for $C_{32}H_{44}O_{11}$ (604.70): C 63.56, H 7.33; found C 63.62, H 7.41%.

19a-β: [α]_D -79.4° (c 1); ¹H-NMR (CDCl₃, 300 MHz, HH-COSY): δ 1.15 (d, 3H, H-6, $J_{6.5}$ 6.6 Hz), 2.16 (s, 3H, CH₃ Ac), 3.28 (ddd, 1H, H-5', $J_{5.4}$ 9.5 Hz, $J_{5.6}$ 2.3 Hz, $J_{5.6}$ 4.8 Hz), 3.39 (s, 3H, CH₃ Me), 3.40 (dd, 1H, H-3', $J_{3.2}$ 3.1 Hz, $J_{3.4}$ 9.0 Hz), 3.47 (dd, 1H, H-4', $J_{4.3} \approx J_{4.5}$ 9.3 Hz), 3.51 (dd, 1H, H-2, $J_{2.1}$ 3.7 Hz, $J_{2.3}$ 10.2 Hz), 3.51, 3.53 (2× s, 6H, 2× CH₃ Me), 3.53 (d, 1H, H-2', $J_{2.3}$ 3.6 Hz), 3.56 (s, 3H, CH₃ Me), 3.74 (dd, 1H, H-6', $J_{6.5}$ 4.8 Hz, $J_{6.6}$ 11.3 Hz), 3.79 (dd, 1H, H-6', $J_{6.5}$ 2.4 Hz, $J_{6.6}$ 11.2 Hz), 3.99 (dq, 1H, H-5, $J_{5.4}$ 1.3 Hz, $J_{5.6}$ 6.6 Hz), 4.24 (dd, 1H, H-3, $J_{3.2}$ 10.0 Hz, $J_{3.4}$ 3.5 Hz), 4.70 (s, 1H, H-1'), 4.84 (d, 1H, H-1, $J_{1.2}$ 3.7 Hz), 5.22 (dd, 1H, H-4, $J_{4.3}$ 3.5 Hz, $J_{4.5}$ 1.3 Hz), 7.22-7.42 (m, 10H, CH Bn); 13 C{ 1 H}-NMR (CDCl₃): δ 15.9 (C-6), 20.7 (CH₃ Ac), 55.2, 59.9, 60.6, 61.3 (4× CH₃ Me), 63.8 (C-5), 69.2 (C-6'), 71.8, 73.5 (2× CH₂ Bn), 70.5, 74.4, 76.2, 76.4, 77.8, 81.9 (CH sugar rings), 98.5 (C-1, ${}^{1}J_{\text{C,H}}$ 172.9 Hz), 98.7 (C-1', ${}^{1}J_{\text{C,H}}$ 153.9 Hz), 127.2, 127.4, 127.6, 127.7, 128.1, 128.2, 128.8 (CH Bn), 138.6 (qC Bn), 170.7 (C=O Ac).

Methyl 4-O-acetyl-3-O-(2,3,6-tri-O-benzyl-4-O-methyl- α -D-mannopyranosyl)-2-O-methyl- α -L-fucopyranoside (19b- α) and Methyl 4-O-acetyl-3-O-(2,3,6-tri-O-benzyl-4-O-methyl- β -D-mannopyranosyl)-2-O-methyl- α -L-fucopyranoside (19b- β) - Condensation of 18(L) with 13 gave, after elution of the silica gel column, the α -linked dimer 19b- α (102 mg, 0.15 mmol) and the β -linked anomer 19b- β (24 mg, 0.03 mmol).

19b-α: $[\alpha]_b$ -28.0° (c 1); ¹H-NMR (CDCl₃, 300 MHz, HH-COSY): δ 1.01 (d, 3H, H-6, $J_{6.5}$ 6.6 Hz), 2.03 (s, 3H, CH₃ Ac), 3.32, 3.39 (2× s, 6H, 2× CH₃ Me), 3.45 (dd, 1H, H-2, $J_{2,1}$ 3.5 Hz, $J_{2,3}$ 10.2 Hz), 3.51 (s, 3H, CH₃ Me), 3.59 (dd, 1H, H-3', $J_{3,2}$ 3.1 Hz, $J_{3,4}$ 9.3 Hz), 3.68 (t, 1H, H-4', $J_{4,3} \approx J_{4,5}$ 9.2 Hz), 3.71 (dd, 1H, H-2', $J_{2,1}$ 2.0 Hz, $J_{2,3}$ 2.8 Hz), 3.76 (AB, 2H, H-6', $J_{6,5}$ 2.3 Hz, $J_{6,5}$ 4.7 Hz), 3.85 (dq, 1H, H-5, $J_{5,4}$ 1.4 Hz, $J_{5,6}$ 6.4 Hz), 4.87 (ddd, 1H, H-5', $J_{5,4}$ 9.6 Hz, $J_{5,6}$ 2.1 Hz, $J_{5,6}$ 4.2 Hz), 4.10 (dd, 1H, H-3, $J_{3,2}$ 10.2 Hz, $J_{3,4}$ 3.5 Hz), 4.54-4.77 (m, 6H, 3× CH₂ Bn), 4.83 (d, 1H, H-1, $J_{1,2}$ 3.6 Hz), 5.15 (dd, 1H, H-4, $J_{4,3}$ 3.6

Hz, J_{45} 1.2 Hz), 5.16 (d, 1H, H-1', $J_{1,2}$ 1.8 Hz), 7.26-7.42 (m, 15H, CH Bn); $^{13}C\{^{1}H\}$ -NMR (CDCl₃): δ 15.7 (C-6), 20.5 (CH₃ Ac), 55.1, 58.6, 60.3 (3× CH₃ Me), 64.5 (C-5), 69.3 (C-6'), 71.7, 71.9, 72.9 (3× CH₂ Bn), 72.1, 72.8, 73.0, 74.1, 76.5, 78.2, 79.0 (CH sugar rings), 97.4, 99.2 (C-1, C-1', $^{1}J_{CH}$ 171.1, 172.9 Hz, respectively), 127.0, 127.3, 127.7, 128.0, 128.1 (CH Bn), 138.1, 138.6, 138.7 (qC Bn), 170.0 (C=O Ac).

Anal. calcd. for C₃₈H₄₈O₁₁ (680.80): C 67.04, H 7.11; found C 67.13, H 7.05%.

19b-β: ¹H-NMR (CDCl₃, 300 MHz, HH-COSY): δ 1.15 (d, 3H, H-6, $J_{6.5}$ 6.6 Hz), 2.05 (s, 3H, CH₃ Ac), 3.31-3.36 (m, 1H, H-5'), 3.40 (s, 3H, CH₃ Me), 3.45 (dd, 1H, H-3', $J_{3.2}$ 2.9 Hz, $J_{2.3}$ 9.4 Hz), 3.53, 3.54 (2× s, 6H, 2× CH₃ Me), 3.67 (t, 1H, H-4', $J_{4.3}$ = $J_{4.5}$ 9.5 Hz), 3.72 (d, 1H, H-2', $J_{2.3}$ 2.8 Hz), 3.80 (d, 1H, H-6', $J_{6.5}$ 4.1 Hz), 3.81 (d, 1H, H-6', $J_{6.5}$ 2.9 Hz), 4.00 (dq, 1H, H-5, $J_{5.4}$ 1.2 Hz, $J_{5.6}$ 6.6 Hz), 4.31 (dd, 1H, H-3, $J_{3.2}$ 10.0 Hz, $J_{3.4}$ 3.5 Hz), 4.53 (s, 1H, H-1'), 4.59 (s, 2H, CH₂ Bn), 4.69, 4.78 (2× AB, 4H, 2× CH₂ Bn), 4.85 (d, 1H, H-1, $J_{1.2}$ 3.7 Hz), 5.24 (dd, 1H, H-4, $J_{4.3}$ 3.6 Hz, $J_{4.5}$ 1.2 Hz), 7.21-7.40 (m, 15H, CH Bn); 13 C{¹H}-NMR (CDCl₃): δ 15.9 (C-6), 20.5 (CH₃ Ac), 55.2, 59.9, 60.7 (3× CH₃ Me), 63.8 (C-5), 69.4 (C-6'), 71.7, 73.6, 73.9 (3× CH₂ Bn), 70.4, 74.1, 75.1, 76.4, 76.5, 82.4 (CH sugar rings), 98.3 (C-1', 11 _{C,H} 153.9 Hz), 98.8 (C-1, 11 _{C,H} 171.4 Hz), 127.3, 127.4, 127.6 127.7, 128.0, 128.1, 128.3, 128.4 (CH Bn), 138.7, 139.0, 139.3 (qC Bn), 171.1 (C=O Ac).

Methyl 4-O-acetyl-3-O-(3,6-di-O-benzyl-2,4-di-O-methyl-D-mannopyranosyl)-2-O-methyl- α -D-fucopyranoside (20a- α) and Methyl 4-O-acetyl-3-O-(3,6-di-O-benzyl-2,4-di-O-methyl- α ,β-D-mannopyranosyl)-2-O-methyl- α -D-fucopyranoside (20a- α ,β) - Condensation of 18(D) with 7 gave, after column chromatography, first the α -linked dimer 20a- α (65.1 mg, 0.11 mmol). Further elution of the column gave a mixture of two anomers (α :β=2:3, 38 mg, 0.06 mmol).

20a-α: $[\alpha]_b$ +138.4° (c 1); ¹H-NMR (CDCl₃, 300 MHz, HH-COSY): δ 1.14 (d, 3H, H-6, $J_{6,5}$ 6.5 Hz), 2.10 (s, 3H, CH₃ Ac), 3.22 (dd, 1H, H-2', $J_{2,1}$ 1.8 Hz, $J_{2,3}$ 2.7 Hz), 3.37 (s, 3H, CH₃ Me), 3.39 (dd, 1H, H-2, $J_{2,3}$ 10.2 Hz), 3.40, 3.47, 3.48 (3× s, 9H, 3× CH₃ Me), 3.63-3.77 (m, 4H, H-3', H-4', H-6'), 3.82 (ddd, 1H, H-5', $J_{5,4}$ 7.8 Hz, $J_{5,6}$ 1.7 Hz, $J_{5,6}$ 3.5 Hz), 3.97 (dq, 1H, H-5, $J_{5,4}$ 1.1 Hz, $J_{5,6}$ 6.5 Hz), 4.16 (dd, 1H, H-3, $J_{3,2}$ 10.2 Hz, $J_{3,4}$ 3.4 Hz), 4.59, 4.70 (2× AB, 4H, 2× CH₂ Bn), 4.83 (d, 1H, H-1, $J_{1,2}$ 3.7 Hz), 5.10 (d, 1H, H-1', $J_{1,2}$ 1.7 Hz), 5.21 (dd, 1H, H-4, $J_{4,3}$ 3.4 Hz, $J_{4,5}$ 1.0 Hz), 7.24-7.41 (m, 10H, CH Bn); ¹³C[¹H}-NMR (CDCl₃): δ 16.0 (C-6), 20.7 (CH₃ Ac), 55.2 (CH₃ 1-O-Me), 58.7, 59.5, 60.4 (3× CH₃ Me), 68.9 (C-6'), 72.0, 73.0 (2× CH₂ Bn), 63.9, 68.9, 69.6, 70.6, 71.3, 76.0, 76.6, 77.8, 78.8 (CH sugar rings), 92.9 (C-1', ¹ $J_{C,H}$ 170.0 Hz), 97.9 (C-1, ¹ $J_{C,H}$ 168.5 Hz), 127.2, 127.4, 127.7, 128.0, 128.1 (CH Bn), 138.6 (qC Bn), 170.5 (C=O Ac).

20b-β: ${}^{13}C\{{}^{1}H\}$ -NMR (CDCl₃): δ 97.6 (C-1, ${}^{1}J_{C,H}$ 167.1 Hz), 102.6 (C-1', ${}^{1}J_{C,H}$ 152.4 Hz). Anal. calcd. for C_{3} , $H_{44}O_{11}$ (604.70): C 63.56, H 7.33; found C 63.45, H 7.45%.

Methyl 4-O-acetyl-3-O-(2,3,6-tri-O-benzyl-4-O-methyl-α-p-mannopyranosyl)-6-deoxy-2-O-methyl-α-p-galactopyranoside (20b-α) and Methyl 4-O-acetyl-3-O-(2,3,6-tri-O-benzyl-4-O-methyl-α,β-p-mannopyranosyl)-6-deoxy-2-O-methyl-α-p-galactopyranoside (20b-α,β) - The two anomers were isolated in a total ratio of $\alpha:\beta=6:1$ after the glycosylation of 18(p) with 13. Elution of the column gave the pure α -linked dimer 20b- α (87 mg, 0.13 mmol). Further elution yielded a mixture of the anomers in a ratio $\alpha:\beta=1:2$ (23 mg, 0.03 mmol).

20b-α: $[\alpha]_b$ +84.8° (c 1); 1 H-NMR (CDCl $_3$, 300 MHz, HH-COSY): δ 1.14 (d, 3H, H-6, $J_{6.5}$ 6.5 Hz), 2.09 (s, 3H, CH $_3$ Ac), 3.36-3.41 (m, 1H, H-2), 3.36, 3.39 (2× s, 6H, 3× CH $_3$ Me), 3.50-3.53 (m, 1H, H-2'), 3.50 (s, 3H, CH $_3$ Me), 3.67 (dd, 1H, H-3', $J_{3.2}$ 2.8 Hz, $J_{3.4}$ 8.8 Hz), 3.73-3.78 (m, 4H, H-4', H-5', H-6'), 3.97 (q, 1H, H-5, $J_{5.6}$ 6.6 Hz), 4.18 (dd, 1H, H-3, $J_{3.2}$ 10.1 Hz, $J_{3.4}$ 3.1 Hz), 4.51, 4.62, 4.70 (3× AB, 6H, 3× CH $_2$ Bn), 4.82 (d, 1H, H-1, $J_{1.2}$ 3.6 Hz), 5.16 (br s, 1H, H-1'), 5.21-5.24 (m, 1H, H-4), 7.21-7.48 (m, 15H, CH Bn); 13 C 1 H}-NMR (CDCl $_3$): δ 16.0 (C-6), 20.6 (CH $_3$ Ac), 55.2 (CH $_3$ 1-*O*-Me), 59.5, 60.5 (2× CH $_3$ Me), 69.0 (C-6'), 71.6 (2×), 73.0 (3× CH $_2$ Bn), 63.9, 69.5, 70.3, 71.4, 74.4, 76.0, 76.6, 78.8 (CH sugar rings), 93.1 (C-1', 1 J_{C,H} 170.0 Hz), 97.9 (C-1, 1 J_{C,H} 170.0 Hz), 127.1, 127.4, 127.7, 128.0 (CH Bn), 138.5 (qC Bn), 170.3 (C=O Ac).

20b- β : ¹³C{¹H}-NMR (CDCl₃): δ 97.6, 103.0 (C-1, C-1').

Anal. calcd. for C₃₉H₄₈O₁₁ (680.80): C 67.04, H 7.11; found C 67.10, H 7.01%.

4-[2-(Benzyloxycarbonylamino)ethyl]phenyl 2,4-di-O-methyl-3-O-{4-O-benzyl-2-O-methyl-3-O-[4-O-acetyl-2-O-methyl-3-O-[4-O-benzyl-2,4-di-O-methyl-α-D-mannopyranosyl]-α-L-rhamnopy

mmol) were dissolved in a mixture of 1,2-dichloroethane-diethyl ether (1/1, v/v, 1 ml). After stirring for 25 min in the presence of activated molecular sieves (4 Å), the reaction mixture was cooled (0°C), and a suspension of NIS (27 mg, 0.12 mmol) and TfOH (1.3 μ l, 15 μ mol) in the same solvent mixture (1.5 ml) was added. Stirring was continued for 15 min. The reaction was quenched with pyridine (0.1 ml), filtered, and diluted with ethyl acetate (15 ml). The organic solution was washed with aq. Na₂S₂O₃ (20%, 10 ml) and aq. NaHCO₃ (10%, 10 ml), dried (MgSO₄), and filtered. The solvents were removed and the residue was purified by silica gel column chromatography (0 \rightarrow 3% acetone in dichloromethane) to give the α -linked tetramer 21- α (80 mg, 63 μ mol) and the β -linked anomer 21- β (26 mg, 21 μ mol).

The glycosylation of 2 with 7 was also performed in diethyl ether at room temperature using the same procedure as described above, while NIS was added as a solid followed by a solution of TfOH in diethyl ether. Column chromatography gave tetramer 21 (112 mg, 88 μ mol), in an anomeric ratio of α : β =10:1, as was evidenced by 1 H-NMR spectroscopy.

21-α: $[\alpha]_{D}$ -49.4° (c 1); ¹H-NMR (CDCl₃, 400 MHz, HH-COSY): δ 1.06 (d, 3H, H-6", $J_{6.5}$ 6.5 Hz), 1.26 (d, 3H, H-6, $J_{6.5}$ 6.1 Hz), 1.33 (d, 3H, H-6', $J_{6.5}$ 6.2 Hz), 2.04 (s, 3H, CH₃ Ac), 2.76 (t, 2H, CH₂ spacer, $J_{H,H}$ 6.8 Hz), 3.20 (s, 3H, CH₃ Me), 3.21 (t, 1H, H-4, $J_{4.3} \approx J_{4.5}$ 9.6 Hz), 3.41 (s, 6H, 2× CH₃ Me), 3.45 (t, 2H, CH₂ spacer, $J_{H,H}$ 6.5 Hz), 3.46 (dd, 1H, H-2", $J_{2.1}$ 4.3 Hz, $J_{2.3}$ 10.9 Hz), 3.48, 3.50 (2× s, 6H, 2× CH₃ Me), 3.51 (t, 1H, H-4', $J_{4.3} \approx J_{4.5}$ 9.9 Hz), 3.53 (s, 3H, CH₃ Me), 3.66 (dq, 1H, H-5, $J_{5.4}$ 9.4 Hz, $J_{5.6}$ 6.4 Hz), 3.69 (dd, 1H, H-2", $J_{2.1}$ 1.9 Hz, $J_{2.3}$ 3.2 Hz), 3.72 (dd, 1H, H-2, $J_{2.1}$ 2.0 Hz, $J_{2.3}$ 3.4 Hz), 3.77 (dd, 1H, H-6", $J_{5.6}$ 4.7 Hz, $J_{6.6}$ 10.8 Hz), 3.41-3.85 (m, 5H, H-2, H-3, H-4, H-5, H-6 all Manp), 3.93 (dq, 1H, H-5", $J_{5.4}$ 9.4 Hz, $J_{5.6}$ 6.3 Hz), 4.00 (dd, 1H, H-3', $J_{3.2}$ 3.2 Hz, $J_{3.4}$ 9.6 Hz), 4.28 (dq, 1H, H-5", $J_{5.4}$ 1.2 Hz, $J_{5.6}$ 6.6 Hz), 4.62, 4.72 (2× AB, 4H, 2× CH₂ Bn), 4.67-4.76 (m, 1H, H-1"), 4.85 (AB, 2H, CH₂ Bn), 5.10 (s, 2H, CH₂ Z), 5.16 (d, 1H, H-1', $J_{1.2}$ 1.7 Hz), 5.18 (d, 1H, H-1", $J_{1.2}$ 2.8 Hz), 5.23 (dd, 1H, H-4", $J_{4.3}$ 3.4 Hz, $J_{4.5}$ 1.2 Hz, $J_{5.6}$ 6.6 Hz), 4.2.2 (2× CH₂ spacer), 7.23-7.42 (m, 20H, arom); 13 C{ 1 H}-NMR (CDCl₃): δ 16.2, 17.8, 18.1 (C-6, C-6", C-6"), 20.6 (CH₃ Ac), 35.2, 42.2 (2× CH₂ spacer), 57.6, 58.3, 58.6, 58.8, 60.4, 61.0 (6× CH₃ Me), 66.5 (CH₂ Z), 69.2 (C-6"), 72.1, 73.1, 75.0 (3× CH₂ Bn), 65.3, 68.6, 68.7, 71.9, 73.2, 73.3, 76.4, 78.3, 78.5, 78.7, 79.2, 79.6, 80.1, 80.5, 81.5, 81.9 (CH sugar rings), 94.8 (C-1, $J_{C,H}$ 170.0 Hz), 98.2, 98.7, 99.4 (C-1', C-1", C-1", $J_{C,H}$ 167.1, 170.0, 167.1 Hz, respectively), 116.4 (CH spacer), 127.2, 127.3, 127.4, 127.6, 128.0, 128.2, 128.4, 129.7 (CH arom), 132.4, 138.6, 139.0, 155.0, 156.2 (qC arom, C=O Z), 170.2 (C=O Ac).

Anal. calcd. for $C_{69}H_{89}NO_{21}$ (1268.47): C 65.33, H 7.07, N 1.10; found C 65.24, H 7.16, N 1.03%.

21-β: ¹H-NMR (CDCl₃, 400 MHz, HH-COSY): δ 1.16 (d, 3H, H-6", $J_{6.5}$ 6.5 Hz), 1.26 (d, 3H, H-6, $J_{6.5}$ 6.2 Hz), 1.29 (d, 3H, H-6', $J_{6.5}$ 6.2 Hz), 2.16 (s, 3H, CH₃ Ac), 2.76 (t, 2H, CH₂ spacer, $J_{\text{H,H}}$ 6.4 Hz), 3.22 (t, 1H, H-4, $J_{4.3} \approx J_{4.5}$ 9.6 Hz), 3.33 (t, 1H, H-4", $J_{4.3} \approx J_{4.5}$ 10.3 Hz), 3.43 (t, 2H, CH₂ spacer, $J_{\text{H,H}}$ 6.7 Hz), 3.30-3.60 (m, 5H, H-2, H-3, H-5, H-6 all Manp), 3.41 (s, 3H, CH₃ Me), 3.45 (t, 1H, H-4', $J_{4.3} \approx J_{4.5}$ 9.7 Hz), 3.47, 3.48, 3.49 (3× s, 9H, 3× CH₃ Me), 3.49 (dd, 1H, H-2") 3.53, 3.54 (2× s, 6H, 2× CH₃ Me), 3.66 (dq, 1H, H-5, $J_{5.4}$ 9.5 Hz, $J_{5.6}$ 6.3 Hz), 3.68-3.71 (m, 2H, H-2, H-2'), 3.91 (dq, 1H, H-5', $J_{5.4}$ 9.4 Hz, $J_{5.6}$ 6.2 Hz), 4.03 (dd, 1H, H-3", $J_{3.2}$ 3.2 Hz, $J_{3.4}$ 9.6 Hz), 4.07 (dd, 1H, H-3, $J_{3.2}$ 3.2 Hz, $J_{3.4}$ 9.7 Hz), 4.34 (dq, 1H, H-5", $J_{5.4}$ 1.2 Hz, $J_{5.6}$ 6.2 Hz), 4.38 (dd, 1H, H-3", $J_{2.3}$ 10.0 Hz, $J_{3.4}$ 3.5 Hz), 4.70 (s, 2H, CH₂ Z), 5.18 (s, 1H, H-1"), 5.19 (d, 1H, H-1', $J_{1.2}$ 1.6 Hz), 5.19 (d, 1H, H-1", $J_{1.2}$ 3.8 Hz), 5.29 (dd, 1H, H-4", $J_{4.3}$ 3.6 Hz, $J_{4.5}$ 0.8 Hz), 5.45 (d, 1H, H-1, $J_{1.2}$ 1.9 Hz), 6.97-7.12 (m, 4H, CH spacer), 7.22-7.44 (m, 20H, CH arom); $J_{3.6}$ 1.80 (CDCl₃): δ 16.3, 17.8, 18.1 (C-6, C-6', C-6"), 20.8 (CH₃ Ac), 35.2, 42.2 (2× CH₂ spacer), 57.7, 58.8, 59.8, 60.7, 61.0, 61.2 (6× CH₃ Me), 66.6 (CH₂ Z), 69.5 (C-6"), 71.9, 73.6, 75.1 (3× CH₂ Bn), 64.5, 68.6, 68.8, 70.6, 74.3, 76.3, 76.4, 77.9, 79.4, 80.1, 80.7, 81.1, 82.0 (CH sugar rings), 95.0 (C-1, $J_{2.H}$ 168.5 Hz), 98.2 (C-1", $J_{2.H}$ 155.3 Hz), 98.2, 100.6 (C-1', C-1", $J_{2.H}$ 166.9, 168.5 Hz, respectively), 116.5 (CH spacer), 127.3, 127.6, 127.7, 127.9, 128.0, 128.2, 128.3, 128.4, 129.7 (CH arom), 132.4, 138.3, 138.6, 139.0, 155.1, 156.2 (qC arom, C=O Z), 170.8 (C=O Ac).

4-(Aminoethyl)phenyl 2,4-di-O-methyl-3-O-{2-O-methyl-3-O-{4-O-acetyl-2-O-methyl-3-O-(2,4-di-O-methyl- α -D-mannopyranosyl)- α -L-rhamnopyranosyl}- α -L-rhamnopyranosyl}- α -L-rhamnopyranosyl}- α -L-rhamnopyranosyl)- α -L-rhamnopyranosyl}- α -L-rhamnopyranosyl) in a mixture of isopropanol-water-acetic acid (10/5/2, v/v/v, 7 ml) was added palladium on carbon (5%). The reaction mixture was stirred for 66 h under a blanket of hydrogen. The reaction mixture was filtered and the filtrate were evaporated to give, after purification of the residual oil by gel-filtration (methanol), the required tetramer 3 (47 mg, 54 μ mol).

¹H-NMR (D₂O, 400 MHz, HH-COSY): δ 1.13 (d, 3H, H-6", $J_{6,5}$ 6.6 Hz), 1.23 (d, 3H, H-6, $J_{6,5}$ 6.3 Hz), 1.33 (d, 3H, H-6', $J_{6,5}$ 6.3 Hz), 2.19 (s, 3H, CH₃ Ac), 2.86 (t, 2H, CH₂ spacer, $J_{R,H}$ 7.3 Hz), 3.09 (t, 2H, CH₂ spacer, $J_{R,H}$ 7.1 Hz), 3.27 (t, 1H, H-4, $J_{4,3}$ $\approx J_{4,5}$ 9.7 Hz), 3.31 (t, 1H, H-4", $J_{4,3}$ $\approx J_{4,5}$ 9.7 Hz), 3.50, 3.52, 3.54 (5× s, 15H, 5× CH₃ Me), 3.56 (dd, 1H, H-2", $J_{2,1}$ 1.6 Hz, $J_{2,3}$ 3.6 Hz), 3.57 (s, 3H, CH₃ Me), 3.59 (t, 1H, H-4', $J_{4,3}$ $\approx J_{4,5}$ 9.7 Hz), 3.65 (m, 1H, H-5"), 3.71 (dd, 1H,

H-2", $J_{2,1}$ 3.7 Hz, $J_{2,3}$ 10.6 Hz), 3.71 (m, 1H, H-6"), 3.74 (dd, 1H, H-2', $J_{2,1}$ 1.6 Hz, $J_{2,3}$ 3.7 Hz), 3.74 (dd, 1H, H-3", $J_{3,2}$ 3.6 Hz, $J_{3,4}$ 10.0 Hz), 3.78 (dq, 1H, H-5, $J_{5,4}$ 9.8 Hz, $J_{5,6}$ 6.5 Hz), 3.83 (dd, 1H, H-6", $J_{6,5}$ 2.1 Hz, $J_{6,6}$ 12.0 Hz), 3.86 (dq, 1H, H-5', $J_{5,4}$ 9.5 Hz, $J_{5,6}$ 6.2 Hz), 3.91 (dd, 1H, H-2, $J_{2,1}$ 1.9 Hz, $J_{2,3}$ 3.4 Hz), 3.98 (dd, 1H, H-3', $J_{3,2}$ 3.3 Hz, $J_{3,4}$ 9.8 Hz), 4.16 (dd, 1H, H-3, $J_{3,2}$ 3.4 Hz, $J_{3,4}$ 9.7 Hz), 4.23 (dd, 1H, H-3", $J_{3,2}$ 3.4 Hz, $J_{3,4}$ 10.4 Hz), 4.29 (dq, 1H, H-5", $J_{5,4}$ 1.1 Hz, $J_{5,6}$ 6.6 Hz), 5.18 (d, 1H, H-1", $J_{1,2}$ 1.6 Hz), 5.25 (d, 1H, H-1', $J_{1,2}$ 1.6 Hz), 5.31 (dd, 1H, H-4", $J_{4,3}$ 3.5 Hz, $J_{4,5}$ 0.9 Hz), 5.44 (d, 1H, H-1", $J_{1,2}$ 3.8 Hz), 5.70 (d, 1H, H-1, $J_{1,2}$ 1.8 Hz), 7.09-7.27 (m, 4H, CH spacer); 13 C{ 11 H}-NMR (D₂O, 100 MHz, CH-COSY): δ 15.9, 17.4, 17.6 (C-6, C-6', C-6"), 20.8 (CH₃ Ac), 34.6, 41.9 (2× CH₂ spacer), 58.4, 58.8, 59.1, 59.4, 61.4 (5× CH₃ Me), 60.6 (C-6"), 66.5 (C-5"), 69.3 (C-5), 70.3 (C-5'), 70.4 (C-5"), 72.1, 72.9, 73.5, 74.1 (4× CH), 77.7 (C-4"), 78.7 (CH), 78.8 (C-3'), 79.3 (C-3), 80.1 (C-2), 81.0 (2× CH), 82.5 (C-4), 95.1 (C-1), 98.9, 99.0 (C-1', C-1"), 99.2 (C-1"), 118.0, 131.0 (CH spacer), 133.3, 154.9 (qC spacer), 174.2 (C=O Ac); MS: [M+H]⁺ for $C_{23}H_{53}NO_{14}$: m/z 864.5.

4-[2-(Benzyloxycarbonylamino)ethyl]phenyl 2,4-di-O-methyl-3-O-{4-O-benzyl-2-O-methyl-3-O-[4-O-benzyl-4-O-methyl- α -D-mannopyranosyl)- α -L-fucopyranosyl]- α -L-rhamnopyranosyl]- α -L-rhamno

Donor 13 and acceptor 2 were also coupled in diethyl ether (4 ml) at room temperature under the agency of NIS and TfOH. NIS was added to a mixture of donor 13, acceptor 2 and activated molecular sieves (4 Å), followed by a solution of TfOH in diethyl ether. An anomeric mixture of tetramer 22 (123 mg, 91 μ mol) was isolated after column chromatography, in a ratio of α : β =12:1 as was established by 1 H-NMR spectroscopy.

22-α: [α]_n -66.2° (c 1); ¹H-NMR (CDCl₃, 400 MHz, HH-COSY): δ 1.05 (d, 3H, H-6", J_{6.5} 6.6 Hz), 1.26 (d, 3H, H-6, J_{6.5} 6.1 Hz), 1.33 (d, 3H, H-6', J_{6.5} 6.2 Hz), 2.03 (s, 3H, CH₃ Ac), 2.76 (t, 2H, CH₂ spacer, J_{HH} 6.9 Hz), 3.11 (s, 3H, CH₃ Me), 3.21 (t, 1H, H-4, J₄₃=J₄₅ 9.6 Hz), 3.40 (s, 3H, CH₃ Me), 3.41 (dd, 1H, H-2", J₂₁ 1.9 Hz, J₂₃ 3.9 Hz), 3.42 (m, 2H, CH₂ spacer), 3.43 (dd, 1H, H-2", J₂₁ 10.3 Hz, J₂₃ 3.5 Hz), 3.51 (s, 6H, 2× CH₃ Me), 3.52 (t, 1H, H-4', J₄₃≈J₄₅ 9.6 Hz), 3.53 (s, 3H, CH₃ Me), 3.60 (dd, 1H, H-3''', $J_{3,2}$ 3.1 Hz, $J_{3,4}$ 9.4 Hz), 3.67 (dq, 1H, H-5, $J_{5,4}$ 9.4 Hz, $J_{5,6}$ 6.2 Hz), 3.68 (dd, 1H, H-2', $J_{2,1}$ 1.6 Hz, $J_{2,3}$ 3.3 Hz, $J_{2,3}$ 3.4 Hz, $J_{3,6}$ 6.2 Hz), 3.68 (dd, 1H, H-2', $J_{2,1}$ 1.6 Hz, $J_{2,3}$ 3.3 Hz, $J_{2,3}$ 3.4 Hz, $J_{2,3}$ 3.5 Hz, $J_{2,3}$ 3.7 Hz, $J_{2,3}$ 3.7 Hz, $J_{2,3}$ 3.7 Hz, $J_{2,3}$ 3.7 Hz, $J_{2,3}$ 3.8 Hz, $J_{2,3}$ 3.8 Hz, $J_{2,3}$ 3.9 H Hz), 3.70 (t, 1H, H-4", J₄₃=J₄₅ 9.6 Hz), 3.72 (dd, 1H, H-2, J₂₁ 1.8 Hz, J₂₃ 3.5 Hz), 3.76 (dd, 1H, H-6", J₆₅ 2.0 Hz, J₆₆ 10.9 Hz), 3.82 (dd, 1H, H-6", J₆₅ 4.8 Hz, J₆₆ 10.9 Hz), 3.89 (ddd, 1H, H-5", J₅₄ 9.8 Hz, J₅₆ 2.0 Hz, J₅₆ 4.7 Hz), 3.94 (dq, 1H, H-5', J₅₄ 9.4 Hz, J₅₆ 6.2 Hz), 4.00 (dd, 1H, H-3', J₃₂ 3.2 Hz, J₃₄ 9.5 Hz), 4.08 (dd, 1H, H-3, J₃₂ 3.2 Hz, J₃₄ 9.6 Hz), 4.25 (dd, 1H, H-3", $J_{3,2}$ 10.3 Hz, $J_{3,4}$ 3.5 Hz), 4.27 (dq, 1H, H-5", $J_{5,4}$ 1.1 Hz, $J_{5,6}$ 6.2 Hz), 4.54-4.75 (m, 7H, 3× CH $_2$ Bn, CH from CH $_2$ Bn), 5.09 (s, 2H, CH₂ Z), 5.17 (m, 1H, CH from CH₂ Bn), 5.17 (d, 1H, H-1", J₁₂ 3.6 Hz), 5.18 (d, 1H, H-1', J₁₂ 1.9 Hz), 5.18 (d, 1H, H-1", J₁₂ 1.9 Hz), 5.23 (dd, 1H, H-4", J₄₃ 3.5 Hz, J₄₅ 1.1 Hz), 5.47 (d, 1H, H-1, J₁₂ 1.9 Hz), 6.95-7.09 (m, 4H, CH spacer), 7.21-7.43 (m, 25H, CH arom); ¹³C{¹H}-NMR (CDCl₃): δ 16.2, 17.8, 18.1 (C-6, C-6', C-6"), 20.6 (CH₃ Ac), 35.2, 42.2 (2× CH₂ spacer), 57.5, 58.2, 58.8, 60.4, 61.0 (5× CH, Me), 66.5 (CH, Z), 69.5 (C-6"), 71.8, 71.9, 73.1, 75.0 (4× CH, Bn), 65.3, 68.6, 68.7, 72.1, 72.9, 73.2, 74.4, 76.6, 78.5, 79.1, 79.2, 79.6, 80.1, 80.5, 81.6, 82.0 (CH sugar rings), 94.9 (C-1, ${}^{1}J_{CH}$ 170.0 Hz), 98.3, 99.4 (C-1', C-1", C-1", ¹J_{CH} 171.5, 167.1 (2x) Hz, respectively), 116.4 (CH spacer), 127.2, 127.3, 127.4, 127.7, 128.1, 128.4, 129.7 (CH arom), 132.4, 138.2, 138.7, 139.2, 155.1, 156.2 (qC arom, C=O Z), 170.1 (C=O Ac).

Anal. calcd. for C₇₅H₉₃NO₂₁ (1344.57): C 67.00, H 6.97, N 1.04; found C 66.88, H 6.90, N 1.15%.

22-β: ¹H-NMR (CDCl₃, 400 MHz, HH-COSY): δ 1.16 (d, 3H, H-6", $J_{6.5}$ 6.5 Hz), 1.26 (d, 3H, H-6, $J_{6.5}$ 6.1 Hz), 1.29 (d, 3H, H-6', $J_{6.5}$ 6.2 Hz), 2.04 (s, 3H, CH₃ Ac), 2.76 (t, 2H, CH₂ spacer, J_{HH} 6.8 Hz), 3.22 (t, 1H, H-4, $J_{43} \approx J_{4.5}$ 9.6 Hz), 3.35 (ddd, 1H, H-5", $J_{5.4}$ 9.7 Hz, $J_{5.6}$ 2.3 Hz, $J_{5.6}$ 4.7 Hz), 3.39 (s, 3H, CH₃ Me), 3.44 (m, 2H, CH₂ spacer), 3.47 (t, 1H, H-4", $J_{43} \approx J_{4.5}$ 9.7 Hz), 3.47 (dd, H-3", $J_{3.2}$ 3.2 Hz, $J_{3.4}$ 10.2 Hz), 3.48, 3.49, 3.50 (3× s, 9H, 3× CH₃ Me), 3.52 (dd, 1H, H-2", $J_{2.1}$ 3.7 Hz, $J_{2.3}$ 10.3 Hz), 3.54 (s, 3H, CH₃ Me), 3.60 (t, 1H, H-4", $J_{4.3} \approx J_{4.5}$ 9.5 Hz), 3.66 (dq, 1H, H-5, $J_{5.6}$ 9.6 Hz, $J_{5.6}$ 6.2 Hz), 3.69-3.70 (m, 1H,

H-2), 3.71-3.72 (m, 1H, H-2'), 3.73 (d, 1H, H-2'', $J_{2,1}$ 3.6 Hz), 3.91 (dq, 1H, H-5', $J_{5,4}$ 9.4 Hz, $J_{5,6}$ 6.2 Hz), 4.03 (dd, 1H, H-3', $J_{3,2}$ 3.2 Hz, $J_{3,4}$ 9.5 Hz), 4.08 (dd, 1H, H-3, $J_{3,2}$ 3.2 Hz, $J_{3,4}$ 9.6 Hz), 4.37 (dq, 1H, H-5", $J_{5,4}$ 1.5 Hz, $J_{5,6}$ 6.4 Hz), 4.44 (dd, 1H, H-3", $J_{3,2}$ 10.1 Hz, $J_{3,4}$ 3.5 Hz), 5.09 (s, 2H, CH₂ Z), 5.19-5.21 (m, 3H, H-1', H-1", H-1"), 5.32 (dd, 1H, H-4", $J_{4,3}$ 3.4 Hz, $J_{4,5}$ 2.1 Hz), 5.45 (d, 1H, H-1, $J_{1,2}$ 1.9 Hz), 6.97-7.10 (m, 4H, CH spacer), 7.22-7.42 (m, 25H, CH arom); 13 C{ 1 H}-NMR (CDCl₃): δ 16.4, 17.8, 18.1 (C-6, C-6"), 20.8 (CH₃ Ac), 35.2, 42.3 (2× CH₂ spacer), 57.7, 58.9, 59.8, 60.8, 61.1 (5× CH₃ Me), 66.6 (CH₂ Z), 69.6 (C-6"), 71.8, 73.6, 75.0, 75.2 (4× CH₂ Bn), 64.6, 68.6, 68.8, 70.4, 75.0, 76.6, 76.7, 79.4, 79.6, 80.1, 80.7, 81.3, 82.0, 85.5 (CH sugar rings), 95.0 (C-1, 1 J_{C,H} 168.5 Hz), 98.0 (C-1", 1 J_{C,H} 161.2 Hz), 98.3, 100.6 (C-1', C-1", 1 J_{C,H} 167.0, 168.5 Hz, respectively), 116.5 (CH spacer), 127.2, 127.3, 127.4, 127.5, 127.6, 127.9, 128.0, 128.1, 128.2, 128.3, 128.5, 129.8 (CH arom), 132.4, 138.5, 138.6, 139.1, 155.2, 156.2 (qC arom, C=O Z), 170.9 (C=O Ac).

4-(Aminoethyl)phenyl 2,4-di-O-methyl-3-O-{2-O-methyl-3-O-{4-O-acetyl-2-O-methyl-3-O-(4-O-methyl- α -D-mannopyranosyl)- α -L-rhamnopyranosyl}- α -L-rhamnopyranosyll}- α -L-rhamnopyranosyll}- α -L-rhamnopyranosyll}- α -L-rhamnopyranosyll}- α -L-rhamnopyranosyl

¹H-NMR (D₂O, 400 MHz, HH-COSY): δ 1.14 (d, 3H, H-6", $J_{6.5}$ 6.5 Hz), 1.26 (d, 3H, H-6, $J_{6.5}$ 6.2 Hz), 1.30 (d, 3H, H-6', $J_{6.5}$ 6.3 Hz), 2.21 (s, 3H, CH₃ Ac), 2.96 (t, 2H, CH₂ spacer, $J_{\text{H,H}}$ 7.3 Hz), 3.25 (t, 2H, CH₂ spacer, $J_{\text{H,H}}$ 7.2 Hz), 3.29 (t, 1H, H-4, $J_{4.3} = J_{4.5}$ 9.4 Hz), 3.41 (t, 1H, H-4", $J_{4.3} = J_{4.5}$ 9.7 Hz), 3.51 (s, 6H, 2× CH₃ Me), 3.51, 3.56, 3.59 (3× s, 9H, 3× CH₃ Me), 3.60 (t, 1H, H-4', $J_{4.3} = J_{4.5}$ 9.7 Hz), 3.71 (dd, 1H, H-2", $J_{2.1}$ 3.6 Hz, $J_{2.3}$ 10.6 Hz), 3.72 (dd, 1H, H-2', $J_{2.1}$ 1.7 Hz, $J_{2.3}$ 3.5 Hz), 3.78 (dq, 1H, H-5, $J_{5.4}$ 9.5 Hz, $J_{5.6}$ 6.3 Hz), 3.70-3.80 (m, 3H, H-3, H-5, H-6 all Manp), 3.85 (dd, 1H, H-6", $J_{6.5}$ 2.9 Hz, $J_{6.6}$ 12.0 Hz), 3.87 (dq, 1H, H-5', $J_{5.4}$ 9.5 Hz, $J_{5.6}$ 6.3 Hz), 3.92 (dd, 1H, H-2, $J_{2.1}$ 2.0 Hz, $J_{2.3}$ 3.0 Hz), 3.95 (dd, 1H, H-2", $J_{2.1}$ 1.5 Hz, $J_{2.3}$ 3.3 Hz), 3.98 (dd, 1H, H-3", $J_{3.2}$ 3.2 Hz, $J_{3.4}$ 9.9 Hz), 4.18 (dd, 1H, H-3, $J_{3.2}$ 3.4 Hz, $J_{3.4}$ 9.7 Hz), 4.22 (dd, 1H, H-3", $J_{3.2}$ 10.4 Hz, $J_{3.4}$ 3.5 Hz), 4.29 (q, 1H, H-5", $J_{5.6}$ 6.6 Hz), 5.06 (d, 1H, H-1", $J_{1.2}$ 1.2 Hz), 5.26 (d, 1H, H-1', $J_{1.2}$ 0.8 Hz), 5.31 (d, 1H, H-4", $J_{4.3}$ 3.5 Hz), 5.43 (d, 1H, H-1", $J_{1.2}$ 3.9 Hz), 5.72 (d, 1H, H-1, $J_{1.2}$ 1.5 Hz), 7.08-7.27 (m, 4H, CH spacer); ¹³C{¹H} NMR (D₂O): δ 15.8, 17.4, 17.5 (C-6, C-6", C-6"), 20.8 (CH₃ Ac), 32.7, 41.3 (2× CH₂ spacer), 58.4, 58.8, 59.1, 60.4, 61.6 (5× CH₃ Me), 61.4 (C-6"), 66.5, 69.3, 70.3, 70.6, 71.0, 72.2, 72.9, 73.2, 74.1, 77.3, 78.6, 78.8, 79.3, 80.1, 81.0, 82.4 (CH sugar rings), 95.0 (C-1), 99.0, 99.2, 102.2 (C-1', C-1", C-1"), 118.3, 131.0 (CH spacer), 132.0, 155.1 (qC spacer), 174.2 (C=O Ac). MS: [M+H]* for $C_{30}H_{61}NO_{19}$: m/z 650.2.

Ethyl 3,6-di-O-benzyl-4-O-methyl-1-thio-D-mannopyranoside (23) - Dibutyltin oxide (2.05 g, 8.2 mmol) was added to a solution of compound 12 (2.44 g, 7.5 mmol) in methanol (45 ml) and the suspension was heated under reflux for 2 h. The clear solution was concentrated and the residue was dried by evaporation with toluene. The stannylidene derivative was dissolved in DMF (80 ml), cesium fluoride (1.50 g, 9.9 mmol) and benzyl bromide (1.3 ml, 10.9 mmol) were added. After stirring for 18 h, DMF was evaporated, and the oily residue was taken up in diethyl ether (50 ml). This solution was washed twice with aq. KF (1 M, 20 ml), once with water (15 ml), dried (MgSO₄), and filtered. The filtrate was concentrated and the crude product was purified by column chromatography (0 \rightarrow 30% ethyl acetate in petroleum ether). Concentration of the appropriate fractions gave compound 23 (2.51 g, 6.0 mmol).

 1 H-NMR (CDCl₃): δ 1.27 (t, 3H, CH₃ SEt, $J_{\text{H,H}}$ 7.5 Hz), 2.60 (ABX, 2H, CH₂ SEt), 3.48 (s, 3H, CH₃ OMe), 3.67 (dd, 1H, H-6, $J_{6,5}$ 2.7 Hz, $J_{6,6}$ 11.2 Hz), 3.54-3.81 (m, 1H, H-3), 3.77 (dd, 1H, H-6, $J_{6,5}$ 4.2 Hz, $J_{6,6}$ 10.8 Hz), 3.89 (t, 1H, H-4, $J_{4,3}$ ≈ $J_{4,5}$ 9.4 Hz), 4.02-4.10 (m, 1H, H-5), 4.05 (dd, 1H, H-2, $J_{2,1}$ 1.7 Hz, $J_{2,3}$ 3.2 Hz), 4.60 (AB, 2H, CH₂ Bn), 4.67 (s, 2H, CH₂ Bn), 5.36 (s, 1H, H-1), 7.28-7.38 (m, 10H, CH Bn); 13 C[1 H}-NMR (CDCl₃): δ 14.4 (CH₃ SEt), 24.4 (CH₂ SEt), 60.2 (CH₃ Me), 68.6 (C-6), 71.2, 72.8 (2× CH₂ Bn), 69.3, 71.0, 75.8, 79.8 (C-2, C-3, C-4, C-5), 83.3 (C-1), 127.0, 127.3, 127.8, 127.9 (CH Bn), 137.5, 137.8 (qC Bn).

Anal. calcd. for C₂₃H₃₀O₅S (418.53): C 66.01, H 7.22; found C 65.87, H 7.14%.

Ethyl 3,6-di-O-benzyl-2-O-chloroacetyl-4-O-methyl-1-thio- α -D-mannopyranoside (24) - To a solution of compound 23 (827 mg, 2.0 mmol) in DMF (20 ml) were added chloroacetic anhydride (673 mg, 4.0 mmol) and aq. NaHCO₃ (360 mg, 4.0 mmol). After stirring for 24 h, the reaction mixture was poured into ice water (20 ml) and extracted with dichloromethane (3× 15 ml). The organic layers were collected, washed with water (15 ml) and aq. NaHCO₃ (10%, 15 ml), dried over (MgSO₄), filtered, and

concentrated to dryness. The residue was purified by silica gel column chromatography. The column was eluted with $0\rightarrow20\%$ ethyl acetate in petroleum ether to afford pure 24 (813 mg, 1.7 mmol).

[α]_D 75.4° (c 1); ¹H-NMR (CDCl₃): δ 1.27 (t, 1H, CH₃ SEt, J_{RH} 7.3 Hz), 2.62 (ABX, 2H, CH₂ SEt), 3.47 (s, 3H, CH₃ Me), 3.60 (t, 1H, H-4, J_{4,3}=J_{4,5} 9.5 Hz), 3.63-3.70 (m, 1H, H-3), 3.78 (d 1H, H-6, J_{6,5} 3.1 Hz), 3.83 (d, 1H, H-6, J_{6,5} 3.8 Hz), 4.02-4.08 (m, 1H, H-5), 4.59, 4.60 (2× AB, 4H, 2× CH₂ Bn), 5.31 (s, 1H, H-1), 5.45 (dd, 1H, H-2, J_{2,1} 1.4 Hz, J_{2,3} 3.0 Hz), 7.29-7.33 (m, 10H, CH Bn); ¹³C{ ¹H}-NMR (CDCl₃): δ 14.3 (CH₃ SEt), 24.8 (CH₂ SEt), 40.3 (CH₂ ClAc), 60.2 (CH₃ Me), 68.3 (C-6), 71.3, 72.7 (2× CH₂ Bn), 71.4, 71.9, 75.5, 77.8 (C-2, C-3, C-4, C-5), 81.3 (C-1), 127.0, 127.2, 127.5, 127.7, 127.8 (CH Bn), 137.1, 137.7 (qC Bn), 166.1 (C=O ClAc).

Anal. calcd. for C₂₅H₃₁O₆SCl (495.04): C 60.66, H 6.31; found C 60.74, H 6.40%.

4-[2-(Benzyloxycarbonylamino)ethyl]phenyl 2,4-di-O-methyl-3-O-{4-O-benzyl-2-O-methyl-3-O-[4-O-acetyl-2-O-methyl-3-O-(3,6-di-O-benzyl-2-O-chloroacetyl-4-O-methyl- α -D-mannopyranosyl)- α -L-fucopyranosyl]- α -L-rhamnopyranosyl}- α -L-rhamnopyranosyl]- α -L-rhamnopyranoside} orthochloroacetate (26) - To a mixture of trimer acceptor 2 (222 mg, 0.25 mmol), mannopyranoside donor 24 (148 mg, 0.30 mmol) and activated molecular sieves (4 Å) in 1,2-dichloroethane-diethyl ether (1/1, ν / ν , 2 ml) was added at 0°C a suspension of NIS (68 mg, 0.30 mmol) and TfOH (3.3 μ l, 37 μ mol) in the same solvent mixture (2 ml). After stirring for 15 min, the reaction was quenched with pyridine, diluted with dichloromethane (10 ml) and filtered. The solution was washed with aq. Na₂S₂O₃ (20%, 7 ml) and aq. NaHCO₃ (10%, 5 ml), dried (MgSO₄), filtered, and concentrated. The residue was purified by column chromatography [ethyl acetate in petroleum ether (0 \rightarrow 50%)]. Elution of the column gave the α -linked tetramer 25- α (123 mg, 93 μ mol) and the 1,2-orthoester linked tetramer 26 (67 mg, 50 μ mol).

25-α: [α]₀ -63.6° (c 1); ¹H-NMR (CDCl₃, 400 MHz, HH-COSY): δ 1.08 (d, 3H, H-6", $J_{6.5}$ 6.5 Hz), 1.26 (d, 3H, H-6, $J_{6.5}$ 6.1 Hz), 1.35 (d, 3H, H-6', $J_{6.5}$ 6.2 Hz), 2.05 (s, 3H, CH₃ Ac), 2.75 (t, 2H, CH₂ spacer, $J_{\text{R,H}}$ 6.8 Hz), 3.22 (t, 1H, H-4, $J_{4.3} \approx J_{4.5}$ 9.6 Hz), 3.31, 3.43 (2× s, 6H, 2× CH₃ Me), 3.40-3.45 (m, 2H, CH₂ spacer), 3.47, 3.49, 3.53 (3× s, 9H, 3× CH₃ Me), 3.48-3.54 (m, 2H, H-2", H-4"), 3.57 (t, 1H, H-4"), $J_{4.3} \approx J_{4.5}$ 9.7 Hz), 3.65-3.72 (m, 5H, H-2, H-5, H-2', H-3"', H-6"'), 3.81 (AB, 1H, H-6"', $J_{6.5}$ 4.0 Hz), 3.89-3.93 (m, 1H, H-5"'), 3.95 (dq, 1H, H-5', $J_{5.4}$ 10.0 Hz, $J_{5.6}$ 6.5 Hz), 4.00 (dd, 1H, H-3', $J_{3.2}$ 3.2 Hz, $J_{3.4}$ 9.5 Hz), 4.09 (dd, 1H, H-3, $J_{3.2}$ 3.1 Hz, $J_{3.4}$ 9.6 Hz), 4.11 (AB, 2H, CH₂ ClAc), 4.25 (dd, 1H, H-3", $J_{3.2}$ 10.2 Hz, $J_{3.4}$ 3.5 Hz), 4.32 (q, 1H, H-5", $J_{5.6}$ 6.8 Hz), 4.62, 4.63, 4.81 (3× AB, 6H, 3× CH₂ Bn), 5.09 (s, 2H, CH₂ Z), 5.14 (d, 1H, H-1"', $J_{1.2}$ 1.3 Hz), 5.19 (d, 1H, H-1", $J_{1.2}$ 4.0 Hz), 5.20 (d, 1H, H-1', $J_{1.2}$ 2.0 Hz), 5.22 (d, 1H, H-4", $J_{4.3}$ 4.0 Hz), 5.39 (dd, 1H, H-2"', $J_{2.1}$ 2.0 Hz, $J_{2.3}$ 3.0 Hz), 5.47 (d, 1H, H-1, $J_{1.2}$ 1.6 Hz), 6.97-7.10 (m, 4H, CH spacer), 7.26-7.41 (m, 20H, CH arom); 13 C(1 H)-NMR (CDCl₃): δ 16.1, 17.7, 18.0 (C-6, C-6', C-6"), 20.5 (CH₃ Ac), 35.0, 42.2 (2× CH₂ spacer), 40.9 (CH₂ ClAc), 57.5, 58.4, 58.7, 60.5, 61.0 (5× CH₃ Me), 66.5 (CH₂ Z) 69.1 (C-6"), 71.7, 73.1, 75.1 (3× CH₂ Bn), 65.2, 68.4, 68.6, 70.6, 71.6, 72.8, 73.1, 75.6, 76.9, 78.4, 79.1, 79.4, 80.0, 80.4, 81.5, 81.8 (CH sugar rings), 94.7 (C-1, 1 J_{C,H} 168.5 Hz), 98.0, 98.8, 99.3 (C-1', C-1", C-1", C-1", I_{C,H} 174.4, 165.6, 167.0 Hz, respectively), 116.3 (CH spacer), 127.3, 127.5, 127.6, 127.7, 128.0, 128.8, 128.3, 129.6 (CH arom), 132.3, 137.7, 138.2, 138.6, 154.9 (QC arom, C=O Z), 166.7 (C=O ClAc), 170.3 (C=O Ac).

Anal. calcd. for C₂₀H₈₈NO₂₂Cl (1330.93): C 63.17, H 6.66, N 1.05; found C 63.10, H 6.78, N 1.13%.

26: [α]_D -64.0° (c 1); ¹H-NMR (CDCl₃, 400 MHz, HH-COSY): δ 1.12 (d, 3H, H-6", $J_{6.5}$ 6.5 Hz), 1.27 (d, 3H, H-6, $J_{6.5}$ 6.3 Hz), 1.33 (d, 3H, H-6', $J_{6.5}$ 6.3 Hz), 2.03 (s, 3H, CH₃ Ac), 2.75 (t, 2H, CH₂ spacer, J_{HH} 6.9 Hz), 3.23 (t, 1H, H-4, $J_{4.3}$ ≈ $J_{4.5}$ 9.6 Hz), 3.30 (s, 3H, CH₃ Me), 3.31 (t, 1H, H-4", $J_{4.3}$ ≈ $J_{4.5}$ 9.3 Hz), 3.42 (q, 2H, CH₂ spacer, J_{HH} 6.5 Hz), 3.47 (s, 3H, CH₃ Me), 3.49 (t, 1H, H-4', $J_{4.3}$ ≈ $J_{4.5}$ 9.6 Hz), 3.50, 3.52 (2× s, 6H, 2× CH₃ Me), 3.52 (dd, 1H, H-2", $J_{2.1}$ 3.4 Hz, $J_{2.3}$ 10.1 Hz), 3.54 (s, 3H, CH₃ Me), 3.63-3.73 (m, 7H, H-2, H-5, H-2', H-3"", H-5"", H-6""), 3.82 (AB, 2H, CH₂ ClAc), 3.95 (dq, 1H, H-5', $J_{5.4}$ 9.40 Hz, $J_{5.6}$ 6.3 Hz), 4.05 (dd, 1H, H-3', $J_{3.2}$ 3.1 Hz, $J_{3.4}$ 9.6 Hz), 4.09 (dd, 1H, H-3, $J_{3.2}$ 3.3 Hz, $J_{3.4}$ 9.7 Hz), 4.32 (dd, 1H, H-3", $J_{3.2}$ 10.4 Hz, $J_{3.4}$ 3.5 Hz), 4.33 (q, 1H, H-5", $J_{5.6}$ 6.0 Hz), 4.59 (t, 1H, H-2"', $J_{2.1}$ ≈ $J_{2.3}$ 3.3 Hz), 4.63, 4.76, 4.83 (3× AB, 6H, 3× CH₂ Bn), 5.09 (s, 2H, CH₂ Z), 5.22 (d, 1H, H-1", $J_{1.2}$ 3.7 Hz), 5.21 (s, 1H, H-1'), 5.28 (d, 1H, H-4", $J_{4.3}$ 3.7 Hz), 5.43 (d, 1H, H-1"', $J_{3.1}$ 3.1 Hz), 5.47 (d, 1H, H-1, $J_{1.2}$ 1.7 Hz), 6.97-7.10 (m, 4H, CH spacer), 7.25-7.43 (m, 20H, CH arom); 13 C{ 1 H}-NMR (CDCl₃, 100 MHz, CH-COSY): δ 16.2 (C-6"), 17.7 (C-6), 18.1 (C-6'), 20.8 (CH₃ Ac), 35.1, 42.2 (2× CH₂ spacer), 44.9 (CH₂ ClAc), 57.5, 58.8, 59.1, 60.5, 61.0 (5× CH₃ Me), 65.1 (C-5"), 66.5 (CH₂ Z), 68.5 (C-5), 68.7 (C-5'), 69.4 (C-6"), 70.4 (C-3"), 71.5, 73.3 (2× CH₂ Bn), 73.5 (C-4"), 74.8 (CH₂ Bn), 75.3 (C-4"), 75.8 (CH), 76.1 (C-2"), 76.4 (CH), 77.3 (CH), 79.2 (CH), 79.6 (C-3), 80.1 (CH), 80.5 (CH), 81.3 (C-3'), 82.0 (C-4'), 94.9 (C-1, 1

171.4, 168.5 Hz, respectively), 116.4 (CH spacer), 121.3 (qC orthoester), 127.4, 127.7, 128.0, 128.2, 128.3, 128.4, 129.7 (CH arom), 132.4, 136.5, 137.9, 138.2, 155.1, 156.2 (qC arom, C=O Z), 170.8 (C=O Ac).

Anal. calcd. for C₇₀H₈₈NO₂₂Cl (1330.93): C 63.17, H 6.66, N 1.05; found C 63.23, H 6.73, N 0.98%.

4-[2-(Benzyloxycarbonylamino)ethyl]phenyl 2,4-di-O-methyl-3-O-{4-O-benzyl-2-O-methyl-3-O-[4-O-acetyl-2-O-methyl-3-O-[4-O-benzyl-4-O-methyl-α-D-mannopyranosyl]-α-L-rhamnopyranos

 $[\alpha]_{\text{b}} \text{-}67.6^{\circ} \text{ (c 1); } ^{13}\text{C}\{^{1}\text{H}\}\text{-NMR (CDCl}_{3}): \delta \ 16.1, 17.7, 18.0 \ (3\times\text{C-6}), 20.6 \ (\text{CH}_{3} \text{ Ac}), 35.2, 42.3 \ (2\times\text{CH}_{2} \text{ spacer}), 57.5, 58.4, 58.8, 60.5, 61.1 \ (5\times\text{CH}_{3} \text{ Me}), 66.6 \ (\text{CH}_{2} \text{ Z}), 69.1 \ (\text{C-6'''}), 71.9, 73.3, 75.0 \ (3\times\text{CH}_{2} \text{ Bn}), 65.2, 68.6, 68.7, 68.8, 71.4, 73.0, 73.3, 75.9, 78.4, 79.2, 79.2, 79.7, 80.1, 80.6, 82.0 \ (\text{CH sugar rings}), 94.9 \ (\text{C-1}), 98.3, 99.6, 101.0 \ (\text{C-1'}, \text{C-1'''}, \text{C-1'''}), 116.5 \ (\text{CH spacer}), 127.2, 127.4, 127.7, 127.9, 128.1, 128.2, 128.4, 128.5, 129.0, 129.8 \ (\text{CH arom}), 132.4, 136.5, 138.2, 138.4, 139.0 \ (\text{qC arom}), 155.1, 156.3 \ (\text{qC arom}, \text{C=O Z}), 170.4 \ (\text{C=O Ac}).$

Anal. calcd. for C₆₈H₉₇NO₂₁ (1254.45): C 65.11, H 6.99, N 1.11; found C 65.25, H 6.92, N 1.18%.

Ethyl 2-O-acetyl-3,6-di-O-benzyl-4-O-methyl-1-thio-D-mannopyranoside (29) - Mannopyranoside 23 (419 mg, 1.0 mmol) was dried by evaporation with pyridine and dissolved in the same solvent (4 ml). Acetic anhydride (0.14 ml, 1.5 mmol) and DMAP (12 mg, 0.1 mmol) were added. After stirring for 1 h, the reaction was quenched with addition of methanol (0.5 ml). The solution was concentrated and the residual acetic acid was removed by repeated evaporation of toluene. The residue was purified by silica gel column chromatography. The column was eluted with ethyl acetate in petroleum ether (0 \rightarrow 10%) to yield after evaporation of the solvent, product 29 (401 mg, 0.9 mmol).

 $\begin{array}{l} [\alpha]_{\rm b} + 79.6^{\rm o} \ (c\ 1); \ ^{\rm l}{\rm H-NMR} \ (CDCl_3); \ \delta\ 1.27 \ (t,\ 3H,\ CH_3\ SEt,\ J_{\rm H,H}\ 7.4\ Hz),\ 2.13 \ (s,\ 3H,\ CH_3\ Ac),\ 2.61 \ (ABX,\ 2H,\ CH_2\ SEt),\ 3.48 \ (s,\ 3H,\ CH_3\ Me),\ 3.63 \ (t,\ 1H,\ H-4,\ J_{4,3}=J_{4,5}\ 9.5\ Hz),\ 3.68 \ (dd,\ 1H,\ H-6,\ J_{6,6}\ 10.8\ Hz,\ J_{6,5}\ 2.0\ Hz),\ 3.72-3.84 \ (m,\ 2H,\ H-3,\ H-6),\ 4.05 \ (ddd,\ 1H,\ H-5,\ J_{5,4}\ 9.6\ Hz,\ J_{5,6}\ 1.8\ Hz,\ J_{5,6}\ 4.0\ Hz),\ 4.59,\ 4.60 \ (2\times AB,\ 4H,\ 2\times CH_2\ Bn),\ 5.29 \ (d,\ 1H,\ H-1,\ J_{1,2}\ 1.3\ Hz),\ 5.38 \ (dd,\ 1H,\ H-2,\ J_{2,1}\ 1.7\ Hz,\ J_{2,3}\ 3.2\ Hz),\ 7.27-7.36 \ (m,\ 10H,\ CH\ Bn);\ ^{13}C\{^{1}H\}-NMR \ (CDCl_3):\ \delta\ 14.6 \ (CH_3\ SEt),\ 20.8 \ (CH_3\ Ac),\ 25.2 \ (CH_2\ SEt),\ 60.6 \ (CH_3\ Me),\ 68.7 \ (C-6),\ 71.5,\ 73.1 \ (2\times CH_2\ Bn),\ 70.3,\ 71.6,\ 75.8,\ 78.2 \ (C-2,\ C-3,\ C-4,\ C-5),\ 82.1 \ (C-1),\ 127.3,\ 127.4,\ 127.5,\ 127.7,\ 128.0,\ 128.2 \ (CH\ Bn),\ 137.6,\ 138.0 \ (qC\ Bn),\ 170.0 \ (C=O\ Ac). \end{array}$

Anal. calcd. for $C_{25}H_{32}O_6S$ (460.59): C 65.19, H 7.00; found C 65.12, H 7.07%.

Ethyl 2-O-benzoyl-3,6-di-O-benzyl-4-O-methyl-1-thio- α -D-mannopyranoside (30) - Compound 23 (423 mg, 1.0 mmol) was dissolved in pyridine (3 ml) and benzoyl chloride (0.17 ml, 1.5 mmol) was added. After stirring for 1 h, the reaction was quenched with water, and the solvents were removed. The residue was taken up in ethyl acetate (10 ml). The solution was washed with water (5 ml) and aq. NaHCO₃ (10%, 5 ml), dried (MgSO₄), and filtered. The organic layer was concentrated and the oily residue was purified by column chromatography. The column was eluted with ethyl acetate in petroleum ether (0 \rightarrow 20%). Concentration of the appropriate fractions gave compound 30 (452 mg, 0.9 mmol).

[α]_b +29.0° (c 1); ¹H-NMR (CDCl₃): δ 1.29 (t, 1H, CH₃ SEt, J_{H,H} 7.4 Hz), 2.64 (ABX, 2H, CH₂ SEt), 3.52 (s, 3H, CH₃ Me), 3.55-4.14 (m, 5H, H-3, H-4, H-5, H-6), 4.64 (CH₂ Bn), 5.40 (s, 1H, H-1), 5.64-5.66 (m, 1H, H-2), 7.25-7.37 (m, 13H, CH arom), 8.02-8.07 (m, 2H, CH Bz); ¹³C{¹H}-NMR (CDCl₃): δ 14.5 (CH₃ SEt), 25.1 (CH₂ SEt), 60.4 (CH₃ Me), 68.7 (C-6), 71.1, 72.9 (2× CH₂ Bn), 70.5, 71.7, 75.7, 78.1 (C-2, C-3, C-4, C-5), 82.1 (C-1), 126.9, 127.0, 127.2, 127.5, 127.8, 127.9, 128.4, 129.4 (CH arom), 129.5 (qC Bz), 132.7 (CH Bz), 137.5, 138.1 (qC Bn), 165.0 (C=O Bz).

Anal. calcd. for C₃₀H₃₄O₆S (522.67): C 78.23, H 7.44; found C 78.16, H 7.36%.

Ethyl 3,6-di-O-benzyl-2-O-(2-dibromomethyl)benzoyl-4-O-methyl-1-thio-α-D-mannopyranoside (31) - 2-Dibromomethyl-benzoyl chloride (406 mg, 1.3 mmol) was added to a solution of compound 23 (419 mg, 1.0 mmol) pyridine. After stirring for 4 h at room temperature, the reaction mixture was quenched with water (1 ml) and diluted with ethyl acetate (10 ml). The solution

was washed with water (7 ml) and aq. NaHCO₃ (10%, 7 ml), dried (MgSO₄), and filtered. The solvents were removed and the residue was purified by column chromatography (0 \rightarrow 20% ethyl acetate in petroleum ether) to give compound 31 (575 mg, 0.83 mmol).

[α]_b +27.2° (c 1); ¹H-NMR (CDCl₃): δ 1.29 (t, 3H, CH₃ SEt, $J_{R,H}$ 7.4 Hz), 2.64 (ABX, 2H, CH₂ SEt), 3.55 (s, 3H, CH₃ Me), 3.73 (dd, 1H, H-6, $^2J_{6,6}$ 10.9 Hz, $J_{6,5}$ 1.9 Hz), 3.87 (dd, 1H, H-6, $^2J_{6,6}$ 10.8 Hz, $J_{6,5}$ 3.8 Hz), 3.87 (t, 1H, H-4, $J_{4,3}$ ≈ $J_{4,5}$ 9.4 Hz), 3.93 (dd, 1H, H-3, $J_{3,2}$ 3.0 Hz, $J_{3,4}$ 9.2 Hz), 4.13 (ddd, 1H, H-5, $J_{5,4}$ 9.1 Hz, $J_{5,6}$ 1.8 Hz, $J_{5,6}$ 3.8 Hz), 4.60, 4.69 (2× AB, 4H, 2× CH₂ Bn), 5.43 (d, 1H, H-1, $J_{1,2}$ 1.5 Hz), 5.57 (dd, 1H, H-2, $J_{2,1}$ 1.8 Hz, $J_{2,3}$ 2.9 Hz), 7.19 (dt, 1H, CH DBMB, $J_{R,H}$ 7.7 Hz, $^4J_{R,H}$ 1.1 Hz), 7.23-7.37 (m, 10H, CH Bn), 7.57 (dt, 1H, CH DBMB, $J_{R,H}$ 7.7 Hz, $^4J_{R,H}$ 1.2 Hz), 7.84 (dd, 1H, CH DBMB, $J_{R,H}$ 8.0 Hz, $^4J_{R,H}$ 1.3 Hz), 7.98 (s, 1H, CHBr₂ DBMB), 8.12 (dt, 1H, CH DBMB, $J_{R,H}$ 8.0 Hz, $^4J_{R,H}$ 1.0 Hz); 13 C{ 1 H}-NMR (CDCl₃): δ 14.7 (CH₃ SEt), 25.3 (CH₂ SEt), 38.3 (CHBr₂ DBMB), 60.7 (CH₃ Me), 68.7 (C-6), 71.9, 73.1 (2× CH₂ Bn), 71.6, 72.0, 75.9, 78.1 (C-2, C-3, C-4, C-5), 81.8 (C-1), 124.5 (qC DBMB), 127.2, 127.2, 127.5, 127.7, 128.0, 128.1, 129.1, 129.9, 131.3, 132.9 (CH arom), 137.2, 137.9 (qC Bn), 142.6 (qC DBMB), 165 (C=0 DBMB).

Anal. calcd. for C₃₁H₃₄O₆SBr₂ (694.50): C 53.61, H 4.93; found C 53.49, H 5.02%.

Methyl 4-O-acetyl-3-O-(3,6-di-O-benzyl-2-O-chloroacetyl-4-O-methyl- α -D-mannopyranosyl)-2-O-methyl- α -L-fucopyranoside (19c- α) and Methyl 4-O-acetyl-3-O-(3,6-di-O-benzyl-2-O-chloroacetyl-4-O-methyl- β -D-mannopyranosyl)-2-O-methyl- α -L-fucopyranoside (19c- β) and 1,2-(3,6-di-O-Benzyl-4-O-methyl- α -D-mannopyranose) 3-(methyl 4-O-acetyl-2-O-methyl- α -L-fucopyranoside) orthochloroacetate (28) - According to the general glycosylation procedure, acceptor 18(L) was coupled with donor 24 to furnish, after purification by column chromatography, the α -anomer 19c- α (63 mg, 0.09 mmol) together with the β -linked dimer 19c- β (10 mg, 0.02 mmol) and the orthoester-linked dimer 28 (57 mg, 0.09 mmol).

This glycosylation was also performed in the presence on 0.3 equiv. of TfOH. After purification of the crude reaction mixture, the individual dimers $19c-\alpha$ (112 mg, 0.17 mmol) and $19c-\beta$ (17 mg, 0.03 mmol) were isolated.

19c-α [α]_b -36.0° (c 1); ¹H-NMR (CDCl₃, 300 MHz, HH-COSY): δ 1.04 (d, 3H, H-6, $J_{6,5}$ 6.6 Hz), 2.05 (s, 3H, CH₃ Ac), 3.39 (s, 3H, CH₃ Me), 3.48 (s, 6H, 2× CH₃ Me), 3.53 (t, 1H, H-4', $J_{4,3} \approx J_{4,5}$ 9.7 Hz), 3.53 (dd, 1H, H-2, $J_{2,1}$ 3.5 Hz, $J_{2,3}$ 10.1 Hz), 3.71 (dd, 1H, H-3', $J_{3,2}$ 3.2 Hz, $J_{3,4}$ 9.3 Hz), 3.71 (AB, 1H, H-6', $J_{6,6}$ 10.8 Hz, $J_{6,5}$ 1.8 Hz), 3.79 (AB, 1H, H-6', $J_{6,6}$ 11.0 Hz, $J_{6,5}$ 4.4 Hz), 3.88 (q, 1H, H-5, $J_{5,6}$ 6.4 Hz), 3.90 (ddd, 1H, H-5', $J_{5,4}$ 9.5 Hz, $J_{5,6}$ 2.0 Hz, $J_{5,6}$ 4.1 Hz), 4.10 (dd, 1H, H-3, $J_{3,2}$ 10.1 Hz, $J_{3,4}$ 3.5 Hz), 4.10 (AB, 2H, CH₂ ClAc), 4.61, 4.62 (2× AB, 4H, 2× CH₂ Bn), 4.85 (d, 1H, H-1, $J_{1,2}$ 3.5 Hz), 5.47 (d, 1H, H-1', $J_{1,2}$ 1.7 Hz), 5.15 (dd, 1H, H-4, $J_{4,3}$ 3.6 Hz, $J_{4,5}$ 1.1 Hz), 5.37 (dd, 1H, H-2', $J_{2,1}$ 1.8 Hz, $J_{2,3}$ 3.3 Hz), 7.23-7.38 (m, 10H, CH Bn); $J_{1,2}$ 1.7 Hz), 7.10 (CPCl₃): δ 15.8 (C-6), 20.6 (CH₃ Ac), 40.9 (CH₂ ClAc), 55.3 (CH₃ 1-*O*-Me), 58.8, 60.6 (2× CH₃ Me), 68.5 (C-6'), 71.8, 73.1 (CH₂ Bn), 64.5, 70.7, 71.9, 72.9, 75.8, 77.0, 78.3 (CH sugar rings), 97.5, 98.8 (C-1, C-1', $J_{1,H}$ 167.0, 174.9 Hz, respectively), 127.3, 127.4, 127.6, 127.8, 128.1, 128.3 (CH Bn), 137.8, 138.5 (qC Bn), 166.7 (C=O ClAc), 170.3 (C=O Ac).

Anal. calcd. for C₃₃H₄₃O₁₂Cl (667.16): C 59.41, H 6.50; found C 59.32, H 6.43%.

19c-β [α]_b -57.8° (c 0.5); ¹H-NMR (CDCl₃, 300 MHz, HH-COSY): δ 1.15 (d, 3H, H-6, $J_{6.5}$ 6.6 Hz), 2.21 (s, 3H, CH₃ Ac), 3.33 (ddd, 1H, H-5', $J_{5.4}$ 9.4 Hz, $J_{5.6}$ 2.7 Hz, $J_{5.6}$ 3.6 Hz), 3.40 (s, 3H, CH₃ Me), 3.49 (t, 1H, H-4', $J_{4.3} \approx J_{4.5}$ 9.6 Hz), 3.49 (dd, 1H, H-2, $J_{2.1}$ 3.8 Hz, $J_{2.3}$ 9.9 Hz), 3.48, 3.51 (2× s, 6H, 2× CH₃ Me), 3.57 (dd, 1H, H-3', $J_{3.2}$ 3.2 Hz, $J_{3.4}$ 9.2 Hz), 3.78 (d, 1H, H-6', $J_{6.5}$ 3.8 Hz), 3.97 (dq, 1H, H-5, $J_{5.4}$ 1.3 Hz, $J_{5.6}$ 6.5 Hz), 4.10 (AB, 2H, CH₂ ClAc), 4.28 (dd, 1H, H-3, $J_{3.2}$ 10.0 Hz, $J_{3.4}$ 3.5 Hz), 4.60 (d, 1H, H-1', $J_{1.2}$ 1.0 Hz), 4.63, 4.66 (2× AB, 4H, 2× CH₂ Bn), 4.82 (d, 1H, H-1, $J_{1.2}$ 3.7 Hz), 5.20 (dd, 1H, H-4, $J_{4.3}$ 3.5 Hz, $J_{4.5}$ 1.4 Hz), 5.44 (dd, 1H, H-2', $J_{2.1}$ 1.0 Hz, $J_{2.3}$ 3.1 Hz), 7.26-7.38 (m, 10H, CH Bn); $J_{3.5}$ 11-NMR (CDCl₃): δ 16.0 (C-6), 20.8 (CH₃ Ac), 40.9 (CH₂ ClAc), 55.3 (CH₃ 1-O-Me), 59.9, 60.9 (2× CH₃ Me), 68.8 (C-6'), 71.6, 73.5 (2× CH₂ Bn), 63.8, 70.0, 73.9, 75.7, 76.1, 76.3, 79.7 (CH sugar rings), 95.2 (C-1', $J_{C.H}$ 158.3 Hz), 98.6 (C-1, $J_{C.H}$ 175.8 Hz), 127.4, 127.6, 127.8, 128.0, 128.2, 128.4 (CH Bn).

28: [α]₀ -67.8° (c 1); ¹H-NMR (CDCl₃, 300 MHz, HH-COSY): δ 1.12 (d, 3H, H-6, $J_{6,5}$ 6.6 Hz), 2.18 (s, 3H, CH₃ Ac), 3.37-3.43 (m, 1H, H-5'), 3.41, 3.50, 3.53 (3× s, 9H, 3× CH₃ Me), 3.64 (dd, 1H, H-2, $J_{2,1}$ 3.6 Hz, $J_{2,3}$ 10.2 Hz), 3.63-3.77 (m, 4H, H-3', H-4', H-6'), 3.80 (AB, 2H, CH₂ ClAc), 3.97 (dq, 1H, H-5, $J_{5,4}$ 1.1 Hz, $J_{5,6}$ 6.5 Hz), 4.21 (dd, 1H, H-3, $J_{3,2}$ 10.2 Hz, $J_{3,4}$ 3.5 Hz), 4.58 (AB, 2H, CH₂ Bn), 4.61 (t, 1H, H-2', $J_{2,1} \approx J_{2,3}$ 3.2 Hz), 4.76 (AB, 2H, CH₂ Bn), 4.88 (d, 1H, H-1, $J_{1,2}$ 3.6 Hz), 5.25 (dd, 1H, H-4, $J_{4,3}$ 3.5 Hz, $J_{4,5}$ 1.2 Hz), 5.47 (d, 1H, H-1', $J_{1,2}$ 2.9 Hz), 7.22-7.45 (m, 10H, CH Bn); 13 C{ 1 H}-NMR (CDCl₃): δ 16.0 (C-6), 20.9 (CH₃ Ac), 45.3 (CH₂ ClAc), 55.2 (CH₃ 1-*O*-Me), 59.4, 60.6 (2× CH₃ Me), 69.2 (C-6'), 71.5, 73.2 (2× CH₂ Bn),

64.5, 70.3, 73.4, 74.7, 75.2, 76.2, 76.3, 77.5 (CH sugar rings), 97.7 (C-1', ${}^{1}J_{C,H}$ 175.8 Hz), 98.0 (C-1, ${}^{1}J_{C,H}$ 170.0 Hz), 121.4 (qC orthoester), 127.3, 127.7, 127.7, 128.2, 128.3 (CH Bn), 137.9, 138.2 (qC Bn), 165.9 (C=O ClAc), 170.7 (C=O Ac).

Anal. calcd. for C₃₃H₄₃O₁₂Cl (667.16): C 59.41, H 6.50; found C 59.48, H 6.61%.

Methyl 4-O-acetyl-3-O-(2-O-acetyl-3,6-di-O-benzyl-4-O-methyl-α-D-mannopyranosyl)-2-O-methyl-α-L-fucopyranoside (19d-α) - Glycosylation of donor 29 with model acceptor 18(L) afforded, after silica gel column chromatography, exclusively the α-linked dimer 19d-α (100 mg, 0.16 mmol).

 $[\alpha]_{\text{b}} - 46.6^{\circ} \text{ (c 1); }^{1}\text{H-NMR (CDCl}_{3}, 300 \text{ MHz, HH-COSY): } \delta 1.03 \text{ (d, 3H, H-6, J}_{6.5} 6.5 \text{ Hz), } 2.04, 2.06 \text{ (2× s, 3H, CH}_{3} \text{ Ac), } 3.38, 3.48, 3.49 \text{ (3× s, 9H, 3× CH}_{3} \text{ Me), } 3.53 \text{ (dd, 1H, H-2, J}_{2,1} 3.6 \text{ Hz, J}_{2,3} 10.2 \text{ Hz), } 3.56 \text{ (t, 1H, H-4', J}_{4,3}=J_{4,5} 9.6 \text{ Hz), } 3.70 \text{ (dd, 1H, H-3', J}_{3,2} 3.4 \text{ Hz, J}_{3,4} 9.4 \text{ Hz), } 3.72 \text{ (dd, 1H, H-6', J}_{6.6} 10.9 \text{ Hz, J}_{6,5} 1.9 \text{ Hz), } 3.80 \text{ (dd, 1H, H-6', J}_{6.6} 11.0 \text{ Hz, J}_{6,5} 4.6 \text{ Hz), } 3.87 \text{ (dq, 1H, H-5, J}_{5,4} 1.3 \text{ Hz, J}_{5,6} 6.5 \text{ Hz), } 3.89 \text{ (ddd, 1H, H-5', J}_{5,4} 9.9 \text{ Hz, J}_{5,6} 1.9 \text{ Hz, J}_{5,6} 4.3 \text{ Hz), } 4.10 \text{ (dd, 1H, H-3, J}_{3,2} 10.1 \text{ Hz, J}_{3,4} 3.6 \text{ Hz), } 4.52-4.73 \text{ (m, 4H, 2× CH}_{2} \text{ Bn), } 4.84 \text{ (d, 1H, H-1, J}_{1,2} 3.5 \text{ Hz), } 5.08 \text{ (d, 1H, H-1', J}_{1,2} 1.8 \text{ Hz), } 5.15 \text{ (dd, 1H, H-4', J}_{4,3} 3.6 \text{ Hz, J}_{4,5} 1.2 \text{ Hz), } 5.34 \text{ (dd, 1H, H-2', J}_{2,1} 1.8 \text{ Hz, J}_{2,3} 3.4 \text{ Hz), } 7.24-7.38 \text{ (m, 10H, CH Bn); }^{13}\text{C}^{\{1\text{H}\}-\text{NMR (CDCl}_{3}\}: } \delta 15.8 \text{ (C-6), } 20.6, 20.9 \text{ (2× CH}_{3} \text{ Ac), } 55.3 \text{ (CH}_{3} 1-O-\text{Me), } 58.9, 60.5 \text{ (2× CH}_{3} \text{ Me), } 69.0 \text{ (C-6'), } 71.5, 73.9 \text{ (2× CH}_{2} \text{ Bn), } 64.5, } 68.8, 71.9, 72.8, 72.9, 75.9, 77.1, 78.4 \text{ (CH sugar rings), } 97.6, 99.1 \text{ (C-1, C-1', $^{1\text{J}}_{\text{C,H}}} 171.4 \text{ Hz, both), } 127.2, 127.4, 127.5, 127.7, } 128.1, 128.2 \text{ (CH Bn), } 138.10, 138.67 \text{ (qC Bn), } 170.1, 170.2 \text{ (2× C=O Ac).} }$

Anal. calcd. for C₃₃H₄₄O₁₂ (632.71): C 62.65, H 7.01; found C 62.74, H 6.91%.

Methyl 4-O-acetyl-3-O-(2-O-benzoyl-3,6-di-O-benzyl-4-O-methyl-α-D-mannopyranosyl)-2-O-methyl-α-L-fucopyranoside (19e-α) - Donor 30 was condensed with acceptor 18(L) and the crude product was purified by column chromatography to give pure $19e-\alpha$ (157 mg, 0.23 mmol).

 $[\alpha]_{\rm b}\ -76.0^{\rm o}\ ({\rm c}\ 1);\ ^{\rm i}\ H-NMR\ (CDCl_3,\ 300\ MHz,\ HH-COSY):\ \delta\ 1.06\ ({\rm d},\ 3H,\ H-6,\ J_{6.5}\ 6.5\ Hz),\ 2.06\ ({\rm s},\ 3H,\ CH_3\ Ac),\ 3.38,\ 3.51,\ 3.53\ (3\times {\rm s},\ 9H,\ 3\times CH_3\ Me),\ 3.57\ ({\rm dd},\ 1H,\ H-2,\ J_{2.1}\ 3.6\ Hz,\ J_{2.3}\ 10.1\ Hz),\ 3.78\ ({\rm t},\ 1H,\ H-4',\ J_{4.3}=^{\rm J}_{4.5}\ 9.3\ Hz),\ 3.81-3.96\ ({\rm m},\ 5H,\ H-5',\ H-5',\ H-6'),\ 4.14\ ({\rm dd},\ 1H,\ H-3,\ J_{3.2}\ 10.1\ Hz,\ J_{3.4}\ 3.5\ Hz),\ 4.68,\ 4.69\ (2\times AB,\ 4H,\ 2\times CH_2\ Bn),\ 4.85\ ({\rm d},\ 1H,\ H-1,\ J_{1.2}\ 3.6\ Hz),\ 5.20\ ({\rm dd},\ 1H,\ H-4,\ J_{4.3}\ 3.5\ Hz,\ J_{4.5}\ 1.3\ Hz),\ 5.21\ ({\rm d},\ 1H,\ H-1',\ J_{1.2}\ 1.8\ Hz),\ 5.58\ ({\rm dd},\ 1H,\ H-2',\ J_{2.1}\ 2.1\ Hz,\ J_{2.3}\ 2.7\ Hz),\ 7.23-7.43\ ({\rm m},\ 13H,\ CH\ arom),\ 8.01-8.06\ ({\rm m},\ 2H,\ CH\ Bz);\ ^{13}C(^{1}H)-NMR\ (CDCl_3):\ \delta\ 15.9\ (C-6),\ 20.6\ (CH_3\ Ac),\ 55.3\ (CH_3\ 1-O-Me),\ 59.0,\ 60.6\ (2\times CH_3\ Me),\ 69.5\ (C-6'),\ 71.3,\ 73.2\ (2\times CH_2\ Bn),\ 64.6,\ 69.1,\ 72.1,\ 73.0,\ 73.1,\ 76.1,\ 77.1,\ 78.4\ (CH\ sugar\ rings),\ 97.6,\ 99.2\ (C-1,\ C-1',\ ^{1}J_{C,H}\ 166.9,\ 174.4\ Hz,\ respectively),\ 127.2,\ 127.4,\ 127.6,\ 128.2,\ 129.8\ (CH\ arom),\ 132.9\ (CH\ Bz),\ 138.1,\ 138.8\ (qC\ Bn),\ 165.5\ (C=O\ Bz),\ 170.3\ (C=O\ Ac).$

Anal. calcd. for C₃₈H₄₆O₁₂ (694.78): C 65.69, H 6.67; found C 65.78, H 6.58%.

Methyl 4-O-acetyl-3-O-(3,6-di-O-benzyl-2-O-(2-dibromomethyl)benzoyl-4-O-methyl- α -D-mannopyranosyl)-2-O-methyl- α -L-fucopyranoside (19f- α) - Glycosylation of 18(L) with 31 gave, after purification by silica gel column chromatography, exclusively 19f- α (204 mg, 0.24 mmol).

[α]_D -42.2° (c 1); ¹H-NMR (CDCl₃, 300 MHz, HH-COSY): δ 1.06 (d, 3H, H-6, $J_{6.5}$ 6.5 Hz), 2.09 (s, 3H, CH₃ Ac), 3.37-3.94 (m, 7H, H-2, H-5, H-3', H-4', H-5', H-6'), 3.40, 3.55, 3.56 (3× s, 9H, 3× CH₃ Me), 4.14 (dd, 1H, H-3, $J_{3.2}$ 10.2 Hz, $J_{3.4}$ 3.5 Hz), 4.62, 4.73 (2× AB, 4H, 2× CH₂ Bn), 4.87 (d, 1H, H-1, $J_{1.2}$ 3.6 Hz), 5.19 (br dd, 1H, H-4), 5.24 (d, 1H, H-1', $J_{1.2}$ 1.6 Hz), 5.49 (dd, 1H, H-2', $J_{2.1}$ 2.0 Hz, $J_{2.3}$ 2.6 Hz), 7.21 (dt, 1H, CH DBMB, $J_{R,H}$ 7.6 Hz, ${}^{4}J_{R,H}$ 0.9 Hz), 7.22-7.39 (m, 10H, CH Bn), 7.54 (dt, 1H, CH DBMB, $J_{R,H}$ 7.8 Hz, ${}^{4}J_{R,H}$ 1.2 Hz), 7.78 (dd, 1H, CH DBMB, $J_{R,H}$ 7.9 Hz, ${}^{4}J_{R,H}$ 1.2 Hz), 7.96 (s, 1H, CHBr₂ DBMB), 8.12 (dt, 1H, CH DBMB, $J_{R,H}$ 8.0 Hz, ${}^{4}J_{R,H}$ 0.8 Hz); ${}^{13}C\{{}^{1}H\}$ -NMR (CDCl₃): δ 15.7 (C-6), 20.4 (CH₃ Ac), 38.2 (CHBr₂ DBMB), 55.1 (CH₃ 1-O-Me), 58.8, 60.5 (2× CH₃ Me), 68.8 (C-6'), 71.8, 72.9 (2× CH₂ Bn), 64.3, 70.3, 71.7, 72.7, 75.9, 76.8, 78.2 (CH sugar rings), 97.4, 98.5 (C-1, C-1', ${}^{1}J_{C,H}$ 168.5, 171.5 Hz, respectively), 124.7 (qC DBMB), 127.0, 127.1, 127.4, 127.5, 128.0, 128.1, 129.2, 129.8, 131.2, 132.8 (CH arom), 137.6, 138.3 (qC Bn), 142.4 (qC DBMB), 165.3 (C=O DBMB), 170.0 (C=O Ac).

Anal. calcd. for C₃₀H₄₆O₁₂Br₂ (866.61): C 54.05, H 5.35; found C 54.14, H 5.48%.

Methyl 4-O-acetyl-3-O-(3,6-di-O-benzyl-4-O-methyl- α -D-mannopyranosyl)-2-O-methyl- α -L-fucopyranoside (19g) - Silver perchlorate (141 mg, 0.68 mmol) and 2,6-lutidine (40 μ l, 0.34 mmol) were added to a solution of disaccharide 19f (98 mg, 0.11 mmol) in acetone-water (1.5 ml, 20/1, ν / ν). After stirring for 30 min, lithium bromide (87 mg, 1 mmol) was added to the

reaction mixture and silver bromide was filtered. The solids were washed with a mixture of acetone-water (2 ml, 20/1, v/v) and morpholine (0.4 ml, 4.6 mmol) was added to the filtrate. The reaction mixture was stirred for 45 min and neutralised with acetic acid. The solution was diluted with diethyl ether (10 ml) and washed with water (8 ml), dried (MgSO₄), filtered and concentrated. The residue was purified by column chromatography (30 \rightarrow 60% ethyl acetate in petroleum ether) to yield compound 19g (56 mg, 0.95 mmol).

[α]_p +27.2° (c 1); ¹H-NMR (CDCl₃, 300 MHz, HH-COSY): δ 1.03 (d, 3H, H-6, $J_{6,5}$ 6.6 Hz), 2.08 (s, 3H, CH₃ Ac), 3.40, 3.44, 3.48 (3× s, 9H, 3× CH₃ Me), 3.52 (dd, 1H, H-2, $J_{2,1}$ 3.2 Hz, $J_{2,3}$ 9.8 Hz), 3.54 (t, 1H, H-4', $J_{4,3} \approx J_{4,5}$ 9.3 Hz), 3.61 (dd, 1H, H-3', $J_{3,2}$ 3.0 Hz, $J_{3,4}$ 9.1 Hz), 3.69-3.75 (m, 2H, H-6'), 3.85-3.90 (m, 2H, H-5, H-5'), 3.98 (dd, 1H, H-2', $J_{2,1}$ 1.7 Hz, $J_{2,3}$ 3.0 Hz), 4.12 (dd, 1H, H-3, $J_{3,2}$ 10.2 Hz, $J_{3,4}$ 3.5 Hz), 4.61 (AB, 4H, 2× CH₂ Bn), 4.70 (s, 2H, CH₂ Bn), 4.85 (d, 1H, H-1, $J_{1,2}$ 3.6 Hz), 5.14 (d, 1H, H-1', $J_{1,2}$ 1.5 Hz), 5.16 (br dd, 1H, H-4), 7.25-7.40 (m, 10H, CH Bn); 13 C{ 1 H}-NMR (CDCl₃): δ 15.8 (C-6), 20.6 (CH₃ Ac), 55.3 (CH₃ 1-O-Me), 58.7, 60.5 (2× CH₃ Me), 69.0 (C-6'), 71.9, 72.9 (2× CH₂ Bn), 64.6, 68.7, 71.5, 72.9, 76.0, 78.2, 79.1 (CH sugar rings), 97.5, 100.8 (C-1, C-1'), 127.3, 127.5, 127.7, 128.1, 128.4 (CH Bn), 138.0, 138.5 (qC Bn), 170.3 (C=O Ac).

Anal. calcd. for C₃₁H₄₂O₁₁ (590.67): C 63.04, H 7.17; found C 62.98, H 7.11%.

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